

European Journal of Cell Biology

24th Annual Meeting of the

German Society for Cell Biology Karlsruhe, March 26th - 30th, 2000

The Role of Calcium Influx on Apical Adhesiveness of Human Uterine Epithelial RL95-2 Cells for Trophoblast-like JAR Cells <u>Michael Thie</u>, Hanna Tinel*, and Hans-Werner Denker

Institut für Anatomie. Universitätsklinikum. D-45122 Essen. * Max-Planck-Institut für molekulare Physiologie. D-44227 Dortmund Implantation of mammalian embryos in the uterus is a regulated event that depends on acquisition of adhesive properties of apical plasma membranes in two epithelia, uterine epithelium and trophoblast. A recent hypothesis postulates that uterine epithelial cells (UECs) can enter a state of receptivity by partially down-regulating their polarized phenotype, thus preparing their free surface for cell-cell interaction with the trophoblast. The aim of experiments presented here was to obtain insight into the signaling mechanisms leading to the formation of stable cell-cell bonds between receptive (adhesive) UECs and trophoblast. Using human uterine epithelial RL95-2 cells (RL) which show adhesiveness for human trophoblast-like JAR cells (JAR) we investigated whether the contact of JAR spheroids to RL monolayers elicits changes of intracellular free calcium ([Ca²⁺]) in RL. [Ca²⁺] was determined by ratio imaging in RL loaded with fluorescent indicators (Oregon Green 488 BABTA-1; Fura Red). Impact of JAR spheroids upon, and movement of spheroids across RL produced transient increase in [Ca²⁴] in RL while JAR spheroids simply resting on RL did not. Prolonged rest of JAR spheroids on RL, however, enabled formation of cell-cell bonds. Disruption of these bonds again produced increase in [Ca²⁺], in RL. In each case, the increase of [Ca2+] in RL was due to Ca2+ influx as it could be blocked not only by removing extracellular Ca2+ but also by the Ca2+ channel inhibitor Ni2*. Correspondingly, cell-cell binding of JAR spheroids to RL was blocked by removal of extracellular Ca²⁺ from the culture medium as well as by pretreatment of RL with Ni²⁺. Collectively, these findings demonstrate that Ca²⁺ influx in uterine RL95-2 cells is required for trophoblast-like JAR cell binding. These data suggest that during embryo implantation signaling of the blastocyst to the uterine epithelium involves Ca2+ influx into UECs and that this, in turn, initiates a cascade leading to the formation of stable cell-cell bonds with the trophoblast.