

Molecular Cell Biology Group

Institute of Cell Biology (Cancer Research), IFZ, Virchowstrasse 173, 45147 Essen

<https://www.uk-essen.de/zellbiologie/forschung/arbeitsgruppe-molekulare-zellbiologie/>

Head: Prof. Dr. rer. nat. Verena Jendrossek (Acting Chair IFZ, Head AG I)

<https://www.uni-due.de/zmb/members/verena-jendrossek.php>

UMESciA project Jendrossek Laboratory

(in collaboration with Dept. of Radiotherapy, Prof. Dr. Martin Stuschke)

Relevance of biological factors to outcome of radio(chemo)therapy or radio(chemo)therapy-immunotherapy combinations in advanced solid human tumors

Background:

Concurrent radiochemotherapy (RCTx) and immune checkpoint inhibitor (ICI) therapy have increased survival rates in advanced non-small cell lung cancer (NSCLC) and advanced squamous cell carcinoma of the head and the neck (HNSCC) (1, 2). Besides their well-known direct cytotoxic action, radiotherapy (RT) and cytotoxic chemotherapy (CTx) reprogram the tumor microenvironment (TME), and augment local and systemic antitumor immune responses under certain conditions, particularly when combined with immunotherapy (IT). Innovative treatment concepts therefore combine ICI therapy with standard radiotherapy (RT) or RCTx in order to achieve synergistic antitumor responses (3) and improve therapy outcome, e.g., in patients with advanced NSCLC (4, 5). Promising results of PD1/PD-L1 based ICI therapy in recurrent or metastatic HNSCC (6-8) have also stimulated the clinical development of ICI in combination with RCTx in the curative setting.

Despite improved multimodal therapy, there is still a high risk of therapy failure by locoregional recurrence or metastasis so that prognosis of patients with advanced solid tumors remains poor. The rationale for combining RT or RCTx with ICI therapy is based on the reciprocal induction of common immune effector mechanisms that are linked to efficacy (9-11), whereas tumor-induced or therapy-induced increase in immuno-suppressive cells or mediators may be causative for the limited success of ICI therapy (12). These variables will also impact the risk of developing therapy-induced immune-related adverse effects (irAE), e.g., in the lung and the heart, upon single or combined treatment (13-15).

So far, reliable biomarkers predicting likelihood of RCTx response or risk of recurrence or adverse effects in individual patients are mostly missing. Emerging biomarkers beyond molecular subtype for NSCLC patients or Human Papilloma Virus (HPV)-status (for oropharyngeal squamous cell carcinoma (16)), several aspects of the immune phenotype (17-19) had prognostic value in HPV-negative HNSCC patient cohorts receiving postoperative RCTx, and may also increase the likelihood for response to ICI therapy (2).

Previous work

Our interest focuses on the contribution of tumor-induced, environment-induced and therapy-induced immune mechanisms to therapy failure (primary vs. secondary resistance) or adverse effects of RT or RCTx without or with additional IT in patients suffering from solid human tumors. We have broad experience in preclinical investigations aimed at a biological optimization of RT including the definition of prognostic biomarkers and the identification of therapeutic targets in HNSCC and NSCLC (20-25). Our previous work indicates that RT-induced senescence and reprogramming of the lung environment contribute to facilitated seeding and growth of circulating tumor cells in previously irradiated lung tissue as well as RT-induced pulmonary fibrosis; targeted inhibition of pathology-associated changes in the host immune environment attenuated RT-induced these unfavorable effects in murine models ((26-31)). RT-induced changes in environmental factors altered recruitment and pathologic differentiation of cells from the myeloid compartment and their regulatory cross-talk with cells from the adaptive immune system, particularly T cells, with impact on efficacy and toxicity of RT/RCTx and ICI therapy, and their combination suggested by others (32) and own unpublished work.

Aims and work Program:

There is high medical need to define prognostic markers indicating probability of response and increased risk of recurrence RT, RCTx and IT, and to design more effective combinatorial treatments that reduce the risk of local or distant failure upon and improve therapy outcome in advanced HNSCC (and NSCLC). The proposed project aims to explore how the tumor-induced and therapy-induced changes in the composition of immune cells and cytokines in the peripheral blood and in the TME impact recruitment, phenotype and function of T cells and myeloid cells and the outcome of RT/RCTx and ICI therapy.

Specific aims are as follows:

i) We will use biomaterial from ongoing studies and already collected conserved or vital samples from our co-clinical **murine** HNSCC and NSCLC models to systematically profile composition and phenotype of cells from the innate and adaptive immune system and of immune mediators before therapy and also record time-dependent changes in the host immune repertoire upon exposure to RT, ICI. or the combination (blood/plasma samples; circulating immune cells; tissue samples). We assume that the detailed evaluation of the host immune environment before and under therapy will provide new mechanistic explanations for response or resistance to RT/RCTx, and RT/RCTx-IT combinations and associated immune escape or adverse effects. These findings will reveal additional molecular or immunological determinants that could be used for the design of rational therapeutic strategies suited to avoid primary resistance, relapse, or adverse effects and thereby to optimize therapies involving RT without or with IT in the future. Assumed immune mechanisms underlying therapy failure may be validated in our preclinical models.

ii) We will use biomaterial and molecular data available from ongoing studies and already collected conserved or vital samples (blood/plasma samples/circulating immune cells) from studies with HNSCC (or NSCLC patients) before, during, or after RCTx, ICI therapy, or the combination, if they can be obtained in collaboration with our clinical partners from the Dept. of Radiation Therapy, the Dept. of Otorhinolaryngology and Head and Neck Surgery, and the Dept. of Pathology, to systematically profile various aspects of the host immune environment.

iii) We will use data obtained in aim 1 and 2 as well as datasets from publicly available studies to associate tumor-specific and therapy-induced host immune repertoires with data on tumor response and therapy outcome. Correlation of the data obtained in aim 1 and aim 2 with outcome data upon RT/RCTx and ICI therapy shall provide a scientific basis to define candidate molecular markers or marker patterns suited to predict likelihood of response or risk of therapy failure or adverse effects upon RT/RCTx without or with ICI therapy for future patient stratification (for bioinformatic analyses: collaboration with Prof. Dr. M. Stuschke, Prof. D. Hoffmann; Dr. F. Farahpour). Promising candidate biomarker identified by these investigations will be validated in patient samples (cooperation with Prof. M. Stuschke; Prof. S. Brandau; Prof. S. Lang).

References

1. L. Ma *et al.*, A current review of dose-escalated radiotherapy in locally advanced non-small cell lung cancer. *Radiol Oncol* **53**, 6-14 (2019).
2. A. K. Hess *et al.*, Characterization of the tumor immune microenvironment and its interference with outcome after concurrent chemoradiation in patients with oropharyngeal carcinomas. *Oncoimmunology* **8**, 1614858 (2019).
3. W. Y. Xia *et al.*, Radiotherapy for non-small cell lung cancer in the immunotherapy era: the opportunity and challenge—a narrative review. *Transl Lung Cancer Res* **9**, 2120-2136 (2020).
4. S. J. Antonia *et al.*, Overall Survival with Durvalumab after Chemoradiotherapy in Stage III NSCLC. *N Engl J Med* **379**, 2342-2350 (2018).
5. W. Theelen *et al.*, Effect of Pembrolizumab After Stereotactic Body Radiotherapy vs Pembrolizumab Alone on Tumor Response in Patients With Advanced Non-Small Cell Lung Cancer: Results of the PEMBRO-RT Phase 2 Randomized Clinical Trial. *JAMA Oncol* **5**, 1276-1282 (2019).

6. B. Burtneß *et al.*, Pembrolizumab alone or with chemotherapy versus cetuximab with chemotherapy for recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-048): a randomised, open-label, phase 3 study. *Lancet* **394**, 1915-1928 (2019).
7. P. Szturz, J. B. Vermorkeñ, Immunotherapy in head and neck cancer: aiming at EXTREME precision. *BMC Med* **15**, 110 (2017).
8. K. J. Harrington *et al.*, Nivolumab versus standard, single-agent therapy of investigator's choice in recurrent or metastatic squamous cell carcinoma of the head and neck (CheckMate 141): health-related quality-of-life results from a randomised, phase 3 trial. *Lancet Oncol* **18**, 1104-1115 (2017).
9. R. R. Weichselbaum, H. Liang, L. Deng, Y. X. Fu, Radiotherapy and immunotherapy: a beneficial liaison? *Nat Rev Clin Oncol* **14**, 365-379 (2017).
10. Y. Wang *et al.*, The Reciprocity between Radiotherapy and Cancer Immunotherapy. *Clin Cancer Res* **25**, 1709-1717 (2019).
11. F. Wirsdorfer, V. Jendrossek, Modeling DNA damage-induced pneumopathy in mice: insight from danger signaling cascades. *Radiat Oncol* **12**, 142 (2017).
12. A. Oweida *et al.*, Resistance to Radiotherapy and PD-L1 Blockade Is Mediated by TIM-3 Upregulation and Regulatory T-Cell Infiltration. *Clin Cancer Res* 10.1158/1078-0432.CCR-18-1038 (2018).
13. L. Michel, T. Rassaf, M. Totzeck, Biomarkers for the detection of apparent and subclinical cancer therapy-related cardiotoxicity. *J Thorac Dis* **10**, S4282-S4295 (2018).
14. J. Naidoo *et al.*, Immune-Related Pneumonitis After Chemoradiotherapy and Subsequent Immune Checkpoint Blockade in Unresectable Stage III Non-Small-Cell Lung Cancer. *Clin Lung Cancer* **21**, e435-e444 (2020).
15. F. Teng, M. Li, J. Yu, Radiation recall pneumonitis induced by PD-1/PD-L1 blockades: mechanisms and therapeutic implications. *BMC Med* **18**, 275 (2020).
16. K. K. Ang *et al.*, Human papillomavirus and survival of patients with oropharyngeal cancer. *N Engl J Med* **363**, 24-35 (2010).
17. P. Balermipas *et al.*, CD8+ tumour-infiltrating lymphocytes in relation to HPV status and clinical outcome in patients with head and neck cancer after postoperative chemoradiotherapy: A multicentre study of the German cancer consortium radiation oncology group (DKTK-ROG). *Int J Cancer* **138**, 171-181 (2016).
18. P. Balermipas *et al.*, The PD-1/PD-L1 axis and human papilloma virus in patients with head and neck cancer after adjuvant chemoradiotherapy: A multicentre study of the German Cancer Consortium Radiation Oncology Group (DKTK-ROG). *Int J Cancer* **141**, 594-603 (2017).
19. P. Balermipas *et al.*, Tumour-infiltrating lymphocytes predict response to definitive chemoradiotherapy in head and neck cancer. *Br J Cancer* **110**, 501-509 (2014).
20. J. Hess *et al.*, Genomic amplification of Fanconi anemia complementation group A (FancA) in head and neck squamous cell carcinoma (HNSCC): Cellular mechanisms of radioresistance and clinical relevance. *Cancer Lett* **386**, 87-99 (2017).
21. U. Schoetz *et al.*, Early senescence and production of senescence-associated cytokines are major determinants of radioresistance in head-and-neck squamous cell carcinoma. *Cell Death Dis* **12**, 1162 (2021).
22. J. Hlouschek, C. Hansel, V. Jendrossek, J. Matschke, The Mitochondrial Citrate Carrier (SLC25A1) Sustains Redox Homeostasis and Mitochondrial Metabolism Supporting Radioresistance of Cancer Cells With Tolerance to Cycling Severe Hypoxia. *Front Oncol* **8**, 170 (2018).
23. J. Hlouschek *et al.*, Targeting SLC25A10 alleviates improved antioxidant capacity and associated radioresistance of cancer cells induced by chronic-cycling hypoxia. *Cancer Lett* **439**, 24-38 (2018).
24. J. Matschke *et al.*, Targeted Inhibition of Glutamine-Dependent Glutathione Metabolism Overcomes Death Resistance Induced by Chronic Cycling Hypoxia. *Antioxid Redox Signal* **25**, 89-107 (2016).

25. J. Matschke, E. Wiebeck, S. Hurst, J. Rudner, V. Jendrossek, Role of SGK1 for fatty acid uptake, cell survival and radioresistance of NCI-H460 lung cancer cells exposed to acute or chronic cycling severe hypoxia. *Radiat Oncol* **11**, 75 (2016).
26. C. Hansel *et al.*, Metformin Protects against Radiation-Induced Acute Effects by Limiting Senescence of Bronchial-Epithelial Cells. *Int J Mol Sci* **22** (2021).
27. A. Wiesemann *et al.*, Inhibition of Radiation-Induced Ccl2 Signaling Protects Lungs from Vascular Dysfunction and Endothelial Cell Loss. *Antioxid Redox Signal* **30**, 213-231 (2019).
28. F. Wirsdorfer *et al.*, Extracellular Adenosine Production by ecto-5'-Nucleotidase (CD73) Enhances Radiation-Induced Lung Fibrosis. *Cancer Res* **76**, 3045-3056 (2016).
29. S. de Leve *et al.*, Loss of CD73 prevents accumulation of alternatively activated macrophages and the formation of profibrotic macrophage clusters in irradiated lungs. *FASEB J* **31**, 2869-2880 (2017).
30. D. Klein *et al.*, Mesenchymal Stem Cell Therapy Protects Lungs from Radiation-Induced Endothelial Cell Loss by Restoring Superoxide Dismutase 1 Expression. *Antioxid Redox Signal* **26**, 563-582 (2017).
31. F. Wirsdorfer *et al.*, Thorax irradiation triggers a local and systemic accumulation of immunosuppressive CD4+ FoxP3+ regulatory T cells. *Radiat Oncol* **9**, 98 (2014).
32. E. Wennerberg *et al.*, Barriers to Radiation-Induced In Situ Tumor Vaccination. *Front Immunol* **8**, 229 (2017).