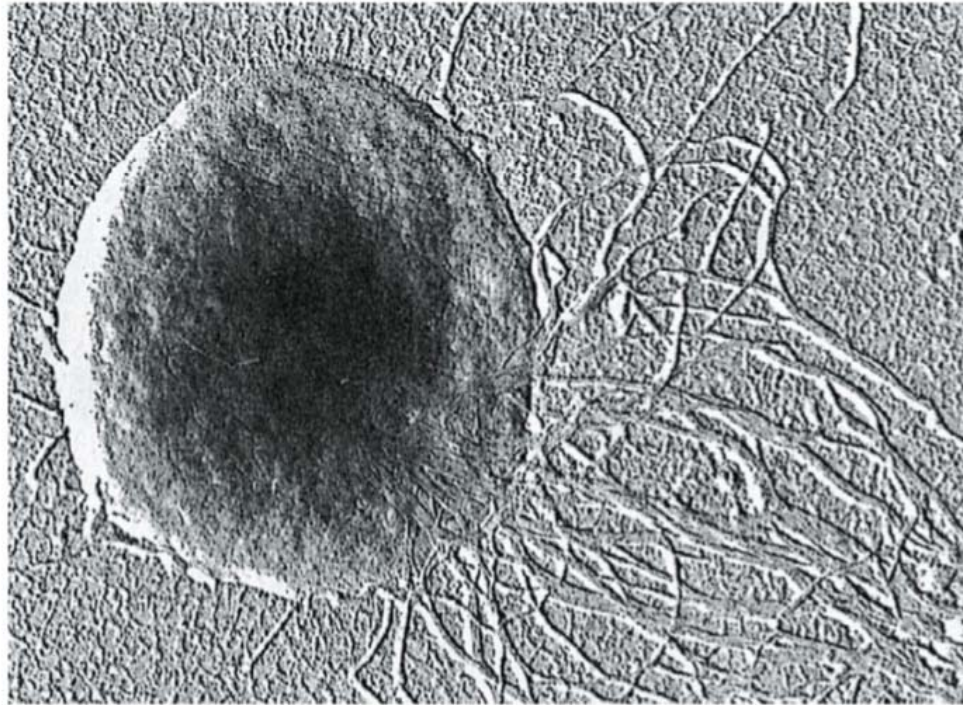


# Microbiology II

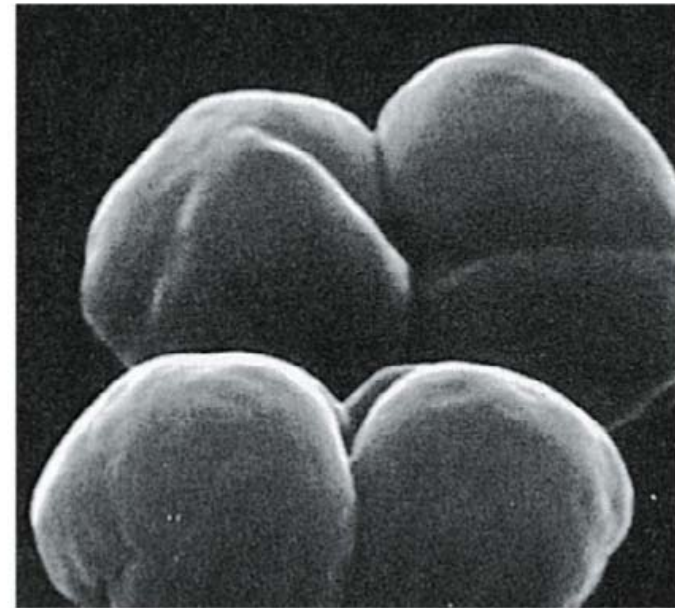
## Methanogenesis



Helmut König und K. O. Stetter

(a)

*Methanococcus jannaschii*



Alexander Zehnder

(d)

*Methanosarcina barkeri*

Christopher Bräsen

# Lecture Plan

17.10. 2017	Mikrobielle Physiologie I - Energetik	Bräsen
24.10. 2017	Mikrobielle Physiologie II – Einige Prinzipien und Mechanismen im zentralen Kohlenstoffmetabolismus	Bräsen
31.10. 2017	Keine Vorlesung	Bräsen
07.11. 2017	Mikrobielle Physiologie III – Nitrat-Atmung	Bräsen
14.11. 2017	Mikrobielle Physiologie IV – Acetogenese und der Acetyl-CoA/Kohlenmonoxid Dehydrogenase-Weg	Bräsen
21.11. 2017	Mikrobielle Physiologie V – Anaerobe Nahrungskette und Methanogenese	Bräsen
28.11. 2017	Mikrobielle Physiologie VI – Sulfate Reduktion	Bräsen
05.12. 2017	Antibiotika (Penicillium notatum)	Meckenstock
12.12. 2017	Mikroorganismen in der Umwelt (Geobacter metallireducens)	Meckenstock
19.12. 2017	Mikrobielles Wachstum (Elusimicrobium minutum)	Meckenstock
09.01. 2018	Mikrobielle Fortbewegung (Thioploca)	Meckenstock
16.01. 2018	Viren (T4)	Meckenstock
23.01. 2018	Geschichte der Mikrobiologie	Meckenstock
30.01. 2018	<b>Wrap up/Ausweichtermin</b>	Meckenstock/Bräsen

# Questions 4

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- What is Acetogenesis, what are the substrates, what the products?
- Name the underlying pathway for CO<sub>2</sub> reduction and the two branches involved?
- What is the key coenzyme?
- What is the key enzyme and which reaction does it catalyze?
- How is energy gained? Which condition is crucial for autotrophic Acetogenesis?
- Outline the steps of the anaerobic food chain! (remember also the different modes of primary fermentations, substrates, products etc.)
- What are the substrates and products of secondary fermentations? What is the key problem and how are these reactions driven?
- What is Syntrophy?

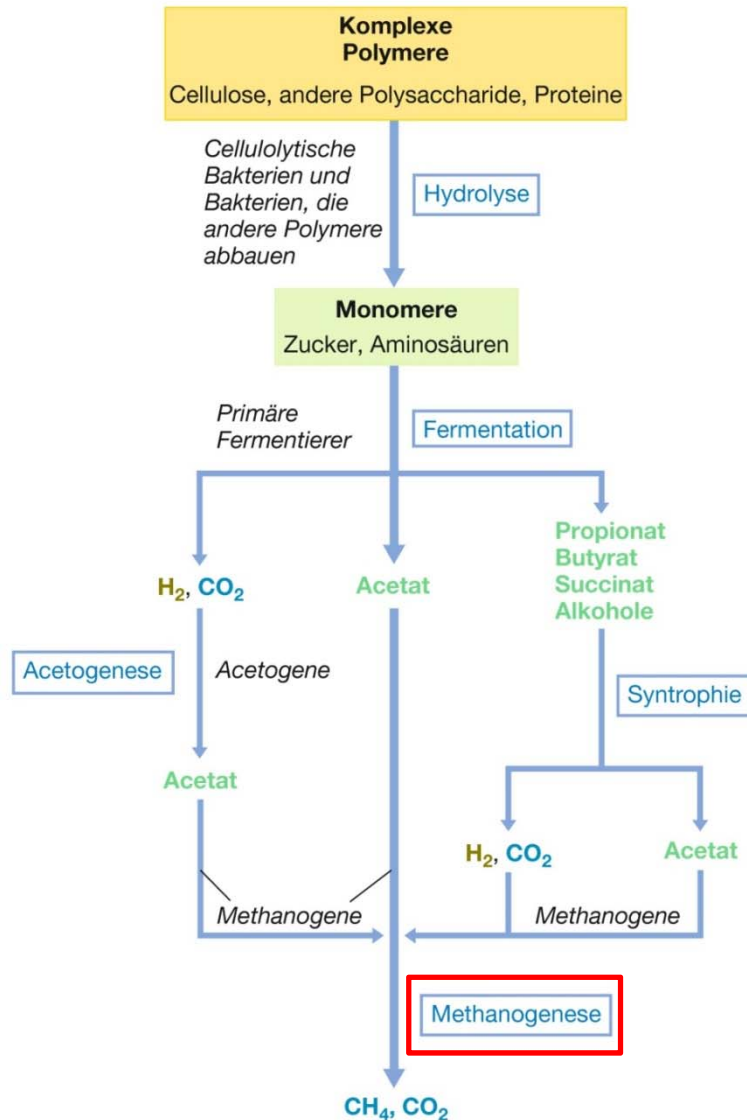
# Fragen 4

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- Was ist Acetogenese, was sind die Substrate, was die Produkte?
- Benenne den zugrunde liegenden Stoffwechselweg zur CO<sub>2</sub> Reduktion und dessen beiden „Zweige“, die daran beteiligt sind?
- Wie heißt das Schlüsselcoenzym dieses Weges?
- Wie heißt das Schlüsselenzym und welche Reaktion katalysiert es?
- Wie hoch ist die Energieausbeute? Welche Bedingung ist für die autotrophe Acetogenese essentiell wichtig?
- Skizzieren Sie die Schritte der anaeroben Nahrungskette! (remember also the different modes of primary fermentations, substrates, products etc.)
- Was sind die Substrate und Produkte sekundärer Fermentationen? Was ist bei diesen Reaktionen das Grundproblem und wie werden sie angetrieben?
- Was ist Syntrophie?

# Anaerobic food chain



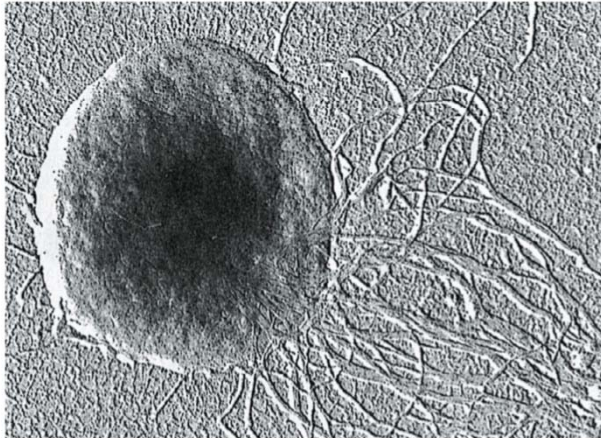
Methane is a major end product of anaerobic biomass degradation only in anoxic environments where the concentrations of sulphate, nitrate, Mn(IV) or Fe(III) are low. In the presence of these electron acceptors, methanogenesis is out-competed by anaerobic respiration, mainly for thermodynamic reasons.

(Thauer RK et al. (2008) Nature Reviews Microbiology 6, 579-591)

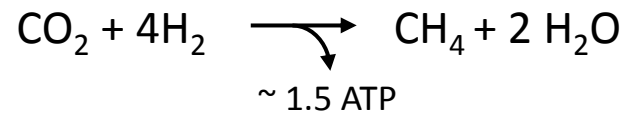
**Abbildung 24.5:** Anoxischer Abbau. Beim anoxischen Abbau kooperieren bei der Umwandlung komplexer organischer Substanzen von  $CH_4$  zu  $CO_2$  verschiedene Gruppen fermentativer Anaerobier. Diese Darstellung trifft auf Lebensräume zu, in denen sulfatreduzierende Bakterien eine untergeordnete Rolle spielen, zum Beispiel in den Sedimenten von Süßwasserseen, Klärschlamm-Bioreaktoren oder dem Pansen.



# Methanogenesis

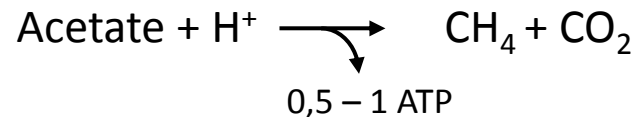


(a) *Methanococcus jannaschii*



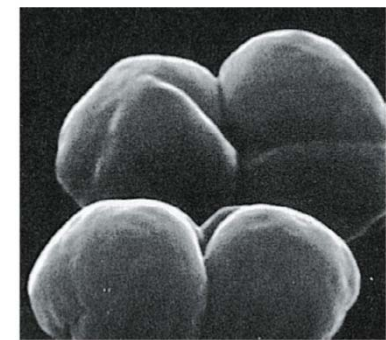
$$\Delta G^{0'} = -131 \text{ kJ/mol}$$

(pH<sub>2</sub> 1 atm)



$$\Delta G^{0'} = -36 \text{ kJ/mol}$$

- Strictly anaerobic Euryarchaea
- Methanobacteriales, Methanopyrales, Methanococcales, Methanomicrobiales, Methanosarcinales
- Most methanogens can grow with CO<sub>2</sub>, formate and H<sub>2</sub>
- Methanosarcinales also with acetate (methylamine, methanol)
- Unusual coenzymes
- 70% of the methane is formed from acetate



(d)

*Methanosarcina barkeri*

# Methanogenesis

## Tabelle 19.4: Habitate der Methanogenen

Anoxische Sedimente: Marschland, Sumpf, Seesedimente, Reisfelder, feuchte Bodenansammlungen

Tierische Verdauungsorgane:

(a) Pansen von Wiederkäuern, zum Beispiel Rinder, Schafe, Elche, Rehe und Kamele

(b) Caecum (Blinddarm) von Tieren mit Caecum wie Pferde und Hasen

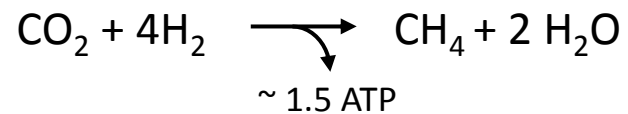
(c) Dickdarm von Tieren mit einem Magen wie bei Menschen, Schweinen und Hunden

(d) Enddarm cellulolytischer Insekten (zum Beispiel Termiten)

Geothermale Quellen von  $H_2 + CO_2$ ; Hydrothermalsysteme

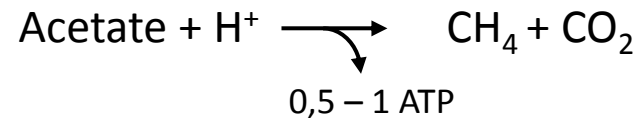
Künstliche Einrichtungen zum biologischen Abbau: Klärschlammanlagen

Endosymbionten verschiedener anaerober Protozoen



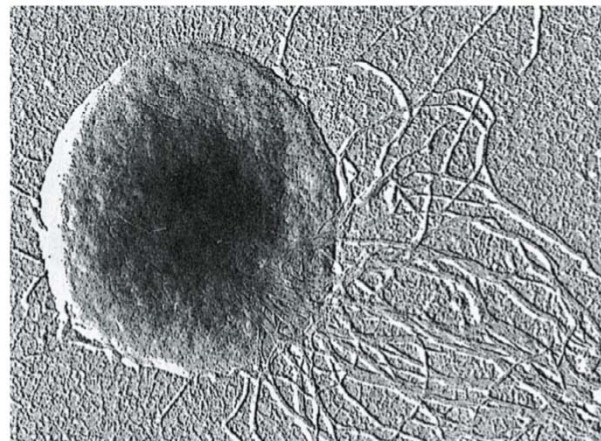
$$\Delta G^{0'} = -131 \text{ kJ/mol}$$

( $p_{H_2} = 1 \text{ atm}$ )



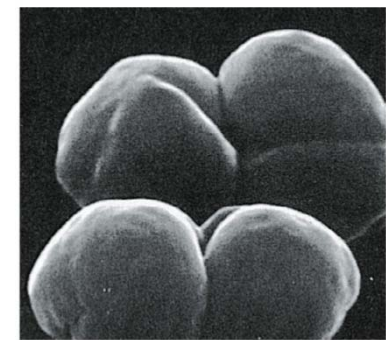
$$\Delta G^{0'} = -36 \text{ kJ/mol}$$

Methane is a very effective green house gas!



(a)

*Methanococcus jannaschii*

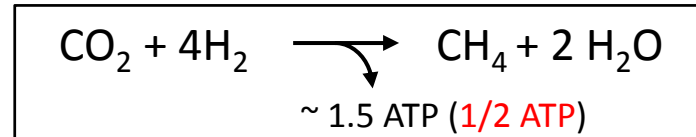


(d)

*Methanosarcina barkeri*

# Methanogenesis from CO<sub>2</sub> and H<sub>2</sub>

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$$\Delta G^{0'} = -131 \text{ kJ/mol}$$

(pH<sub>2</sub> 1 atm)

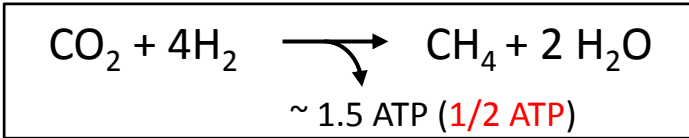
$$\begin{aligned}\Delta G' &= -131 \text{ kJ/mol} + 5.7 \text{ kJ/mol} \times \lg \left( \frac{[\text{P1}]^a [\text{P2}]^b}{[\text{S1}]^c [\mathbf{10^{-5}}]^4} \right) \\ &= -131 \text{ kJ/mol} + 5.7 \text{ kJ/mol} \times 20 \\ &= \mathbf{\sim -17 \text{ kJ/mol (pH}_2 \mathbf{10^{-5} \text{ atm)} \rightarrow \sim 1/3 \text{ ATP}}}\end{aligned}$$

- „Carbonate respiration“
- In natural habitats pH<sub>2</sub> is as low as 10<sup>-5</sup> atm due to methanogenic activity
- This is the lower limit for (autotrophic) methanogenesis
- CO<sub>2</sub> fixation via the rCODH/ACS pathway (=reductive acetyl-CoA pathway)
- Allows however for syntrophic H<sub>2</sub> production



# Methanogenesis from CO<sub>2</sub> and H<sub>2</sub>

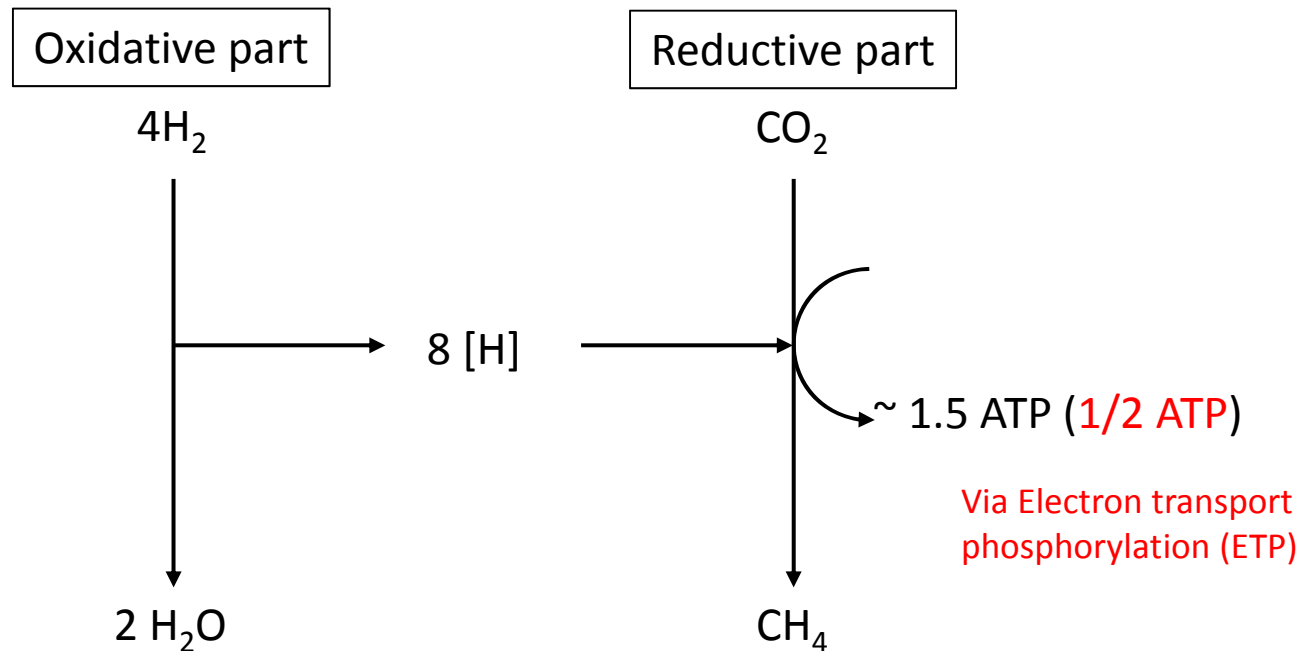
chemolithoautotrophic



$$\Delta G^{\circ'} = -131 \text{ kJ/mol (pH}_2 \text{ 1 atm)}$$

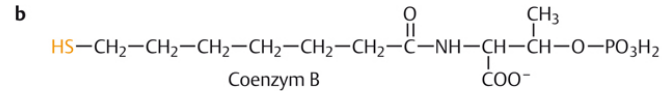
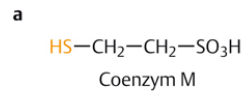
$$\Delta G' = \sim -17 \text{ kJ/mol (pH}_2 \text{ } 10^{-5} \text{ atm)}$$

$$\rightarrow \sim 1/3 \text{ ATP}$$

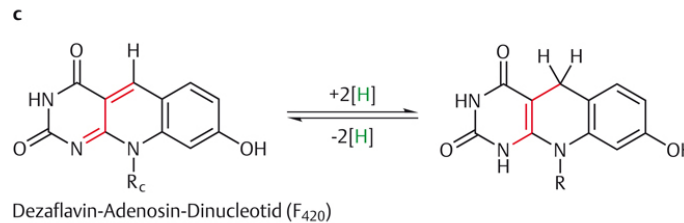


# Methanogenesis

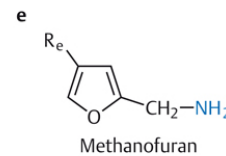
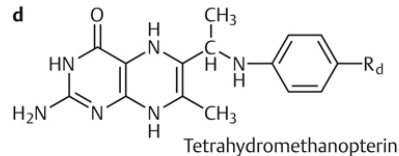
Electron transfer +  
Energy generation



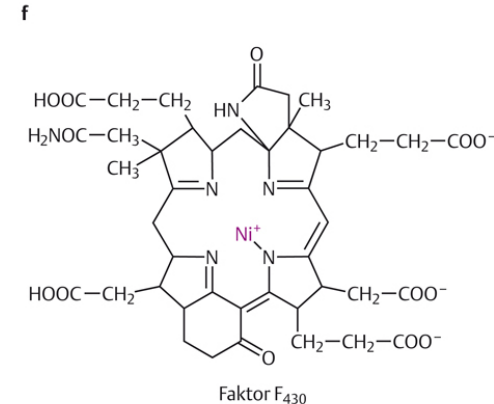
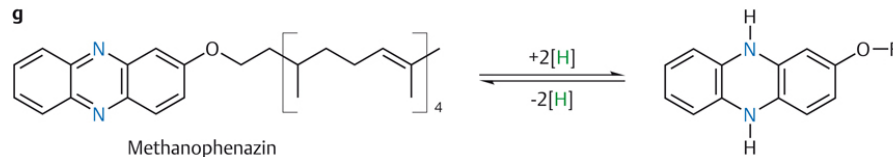
Electron transfer



C1 transfer



Quinone derivative  
Electron transport

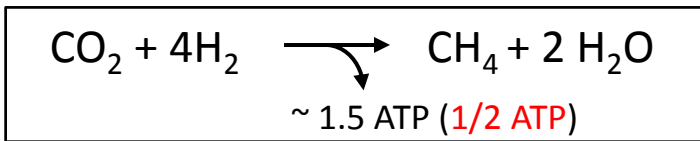


Methyl  
group  
reduction

Georg Thieme Verlag, Stuttgart  
Fuchs et al.: Allgemeine Mikrobiologie, 8. Auflage · 2006

- Strictly anaerobic Euryarchaea
- Methanobacteriales Methanopyrales, Methanococcales, Methanomicrobiales, Methanosarcinales
- Most methanogens can grow with CO<sub>2</sub>, formate and H<sub>2</sub>
- Methanosarcinales also with acetate (methylamine, methanol)
- Unusual coenzymes
- F<sub>420</sub> absorbs light at 420 nm and fluoresce blue-green

# Methanogenesis from CO<sub>2</sub> and H<sub>2</sub>



$\Delta G^{\circ} = -131 \text{ kJ/mol}$ , (pH<sub>2</sub> 1 atm)

$\Delta G' = \sim -17 \text{ kJ/mol}$  (pH<sub>2</sub> 10<sup>-5</sup> atm)

Methanosarcinales with cytochromes and Methanophenazin!

→ higher growth yield

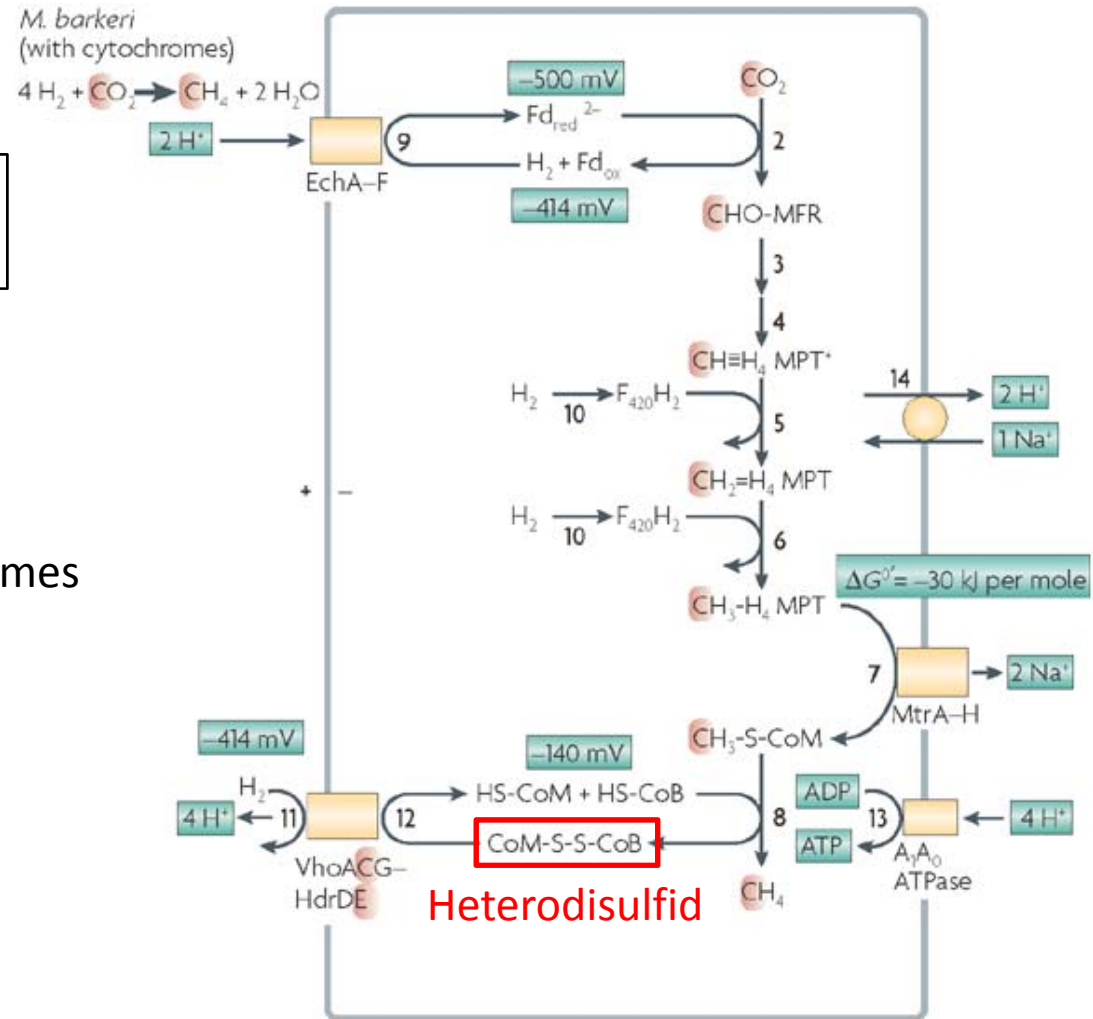
→ higher H<sub>2</sub> threshold

MFR = Methanofuran

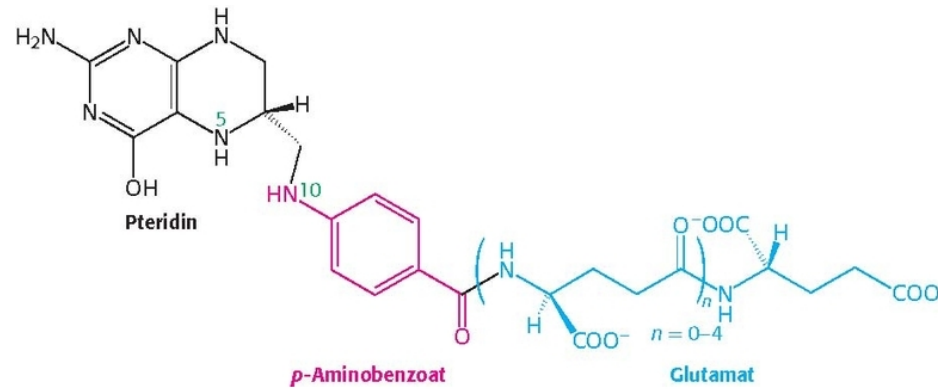
H<sub>4</sub>MPT = Tetrahydromethanopterin

CoM-SH = Coenzym M

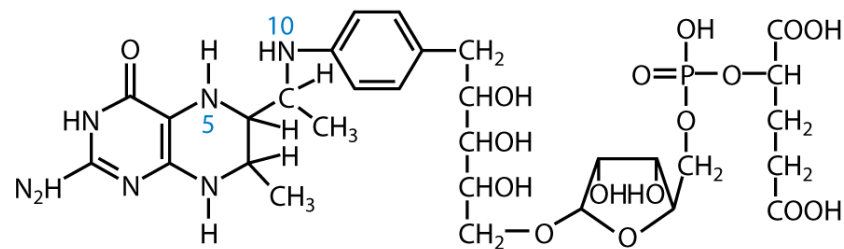
CoB-SH = Coenzym B



# Tetrahydrofolate - Tetrahydromethanopterin

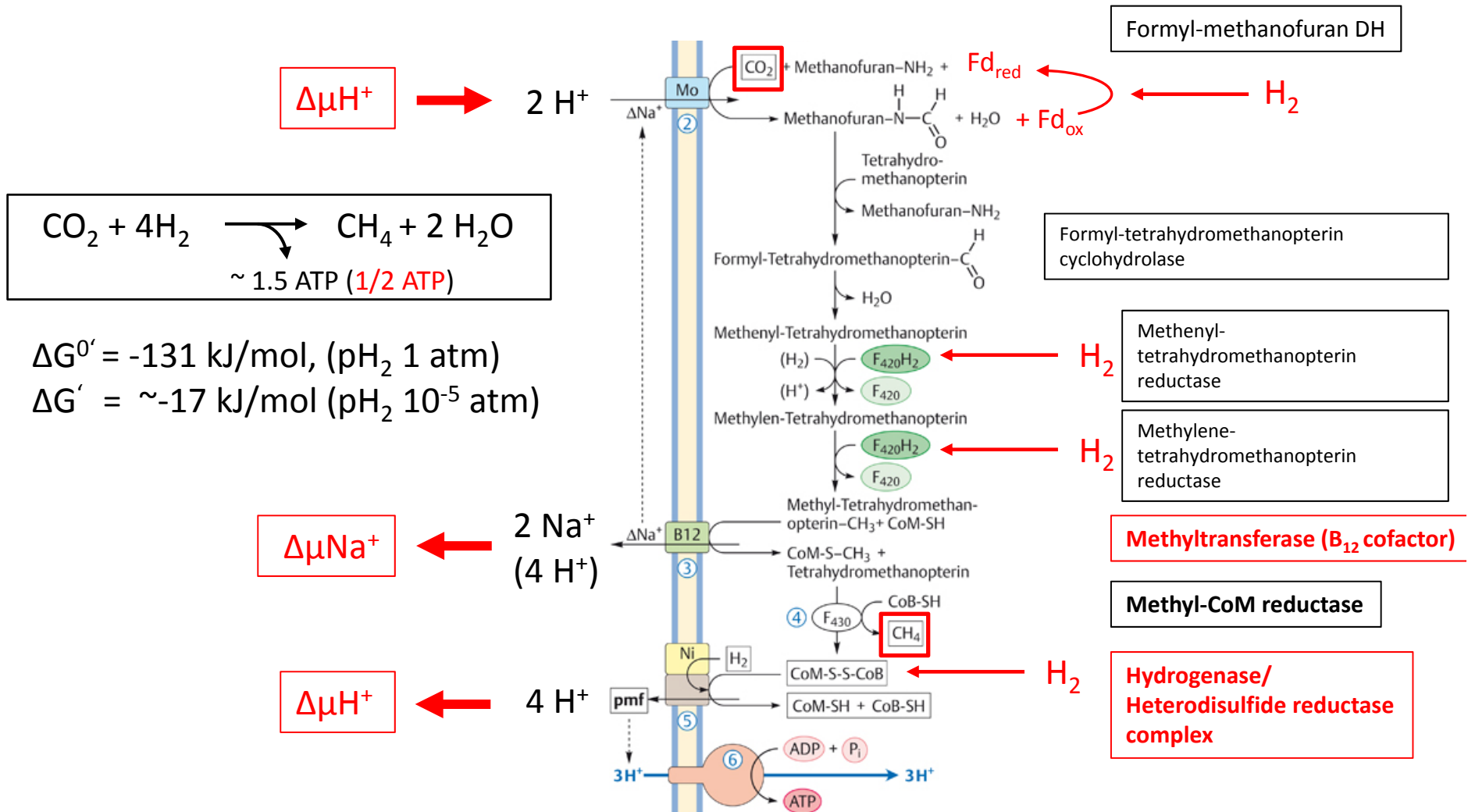


Aus: Berg, Stryer, Tymoczko, Gatto: *Biochemie*, Springer Spektrum © Springer-Verlag Berlin Heidelberg 2013



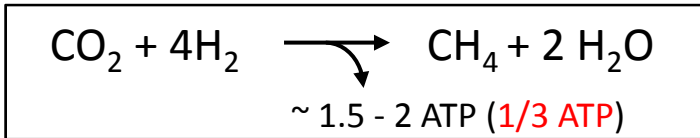
**5,6,7,8-Tetrahydromethanopterin ( $\text{H}_4\text{MPT}$ )**

# Methanogenesis from CO<sub>2</sub> and H<sub>2</sub>





# Methanogenesis from CO<sub>2</sub> and H<sub>2</sub>



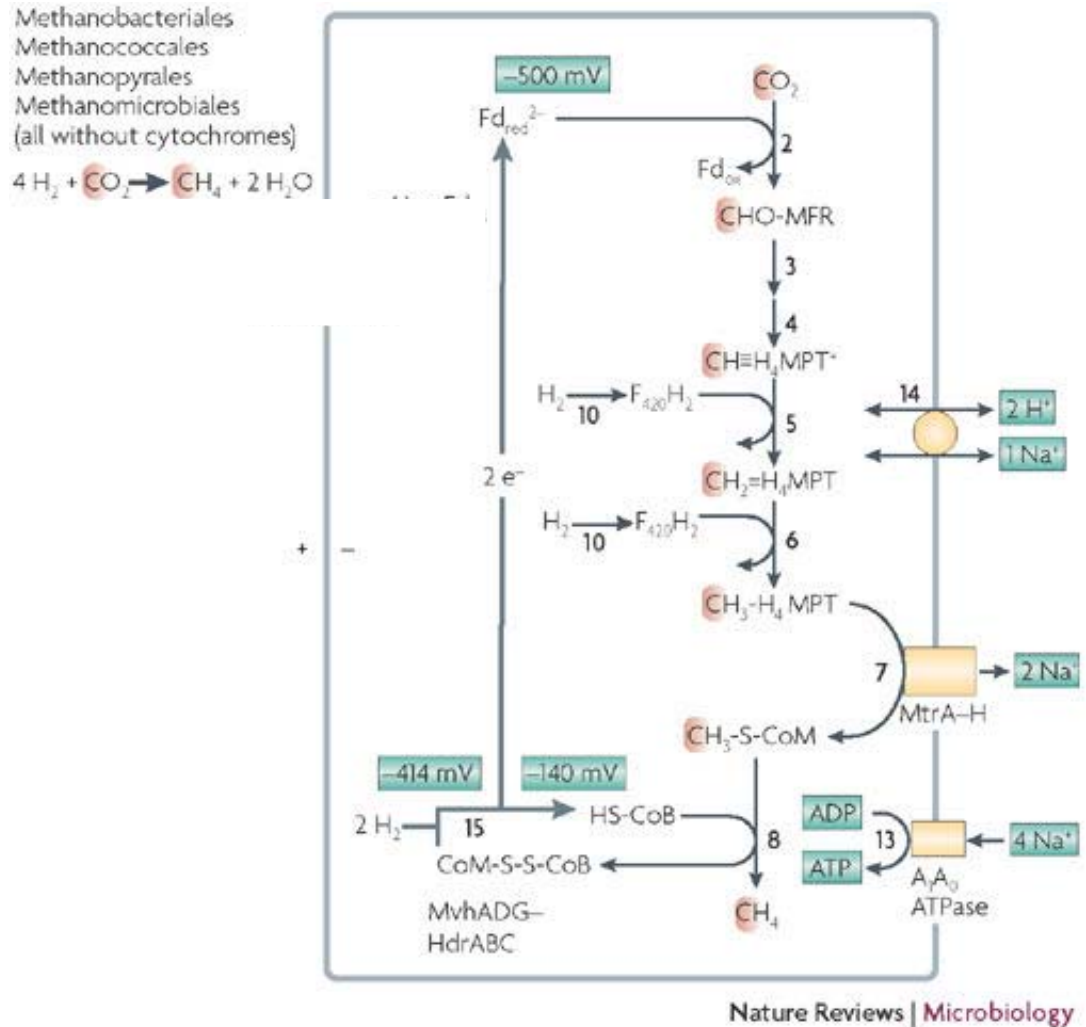
$\Delta G^{\circ} = -131 \text{ kJ/mol}$ , (pH<sub>2</sub> 1 atm)

$\Delta G' = \sim -17 \text{ kJ/mol}$  (pH<sub>2</sub> 10<sup>-5</sup> atm)

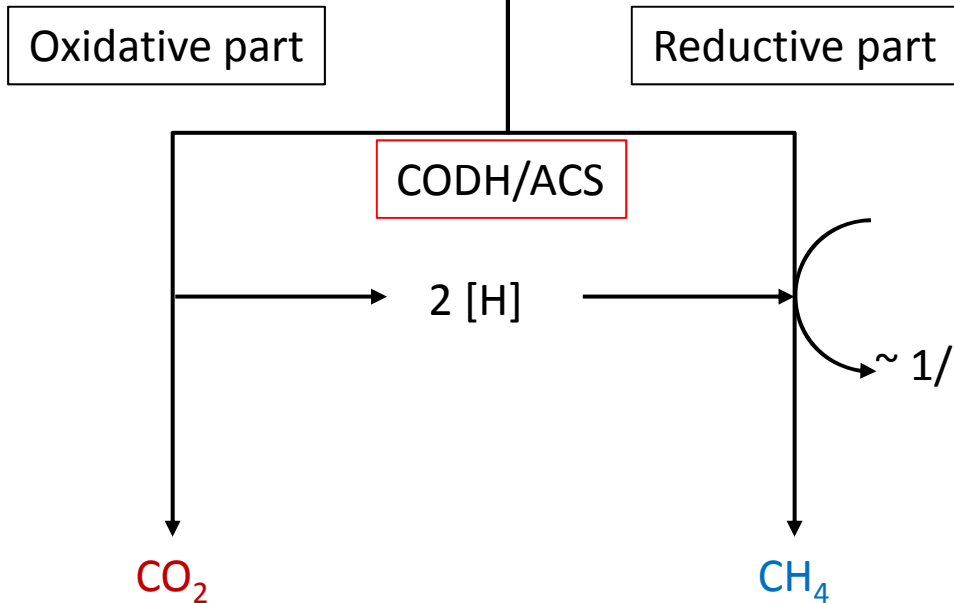
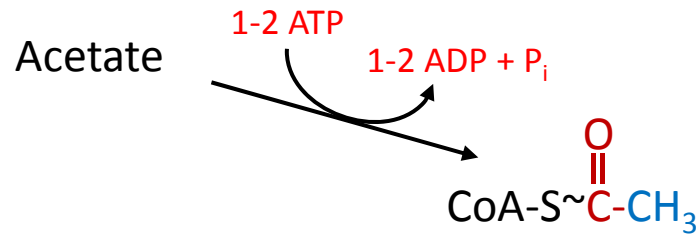
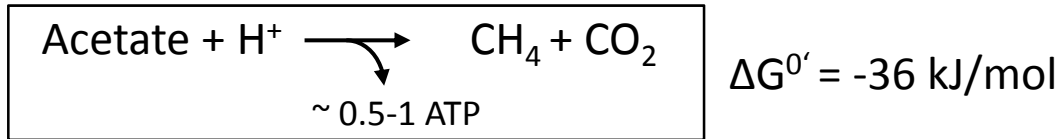
Methanobacteriales,  
Methanococcales, Methanopyrales  
and Methanomicrobiales without  
cytochromes and Methanophenazin!

→ Lower growth yield

→ Lower H<sub>2</sub> threshold



# Methanogenesis from acetate



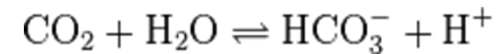
Disproportionation:

- Oxidation of the carboxyl group to  $\text{CO}_2$
- Reduction of the methyl group to  $\text{CH}_4$

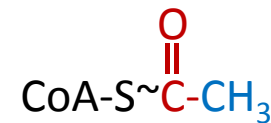
Transport of 7  $\text{H}^+$  ( $\text{Na}^+$ )/acetate  
 $\rightarrow$  corresponding to  $\sim 2 \text{ ATP}$

But: Activation of acetate to acetyl-CoA  
 requires at least 1 ATP

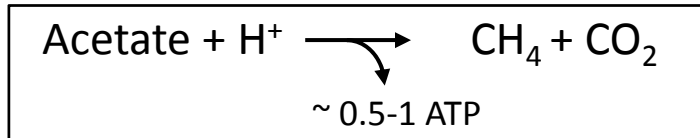
Carbonic anhydrase:



Via Electron transport  
 phosphorylation (ETP)



# Methanogenesis from acetate



Disproportionation:  $\Delta G^{\circ} = -36 \text{ kJ/mol}$

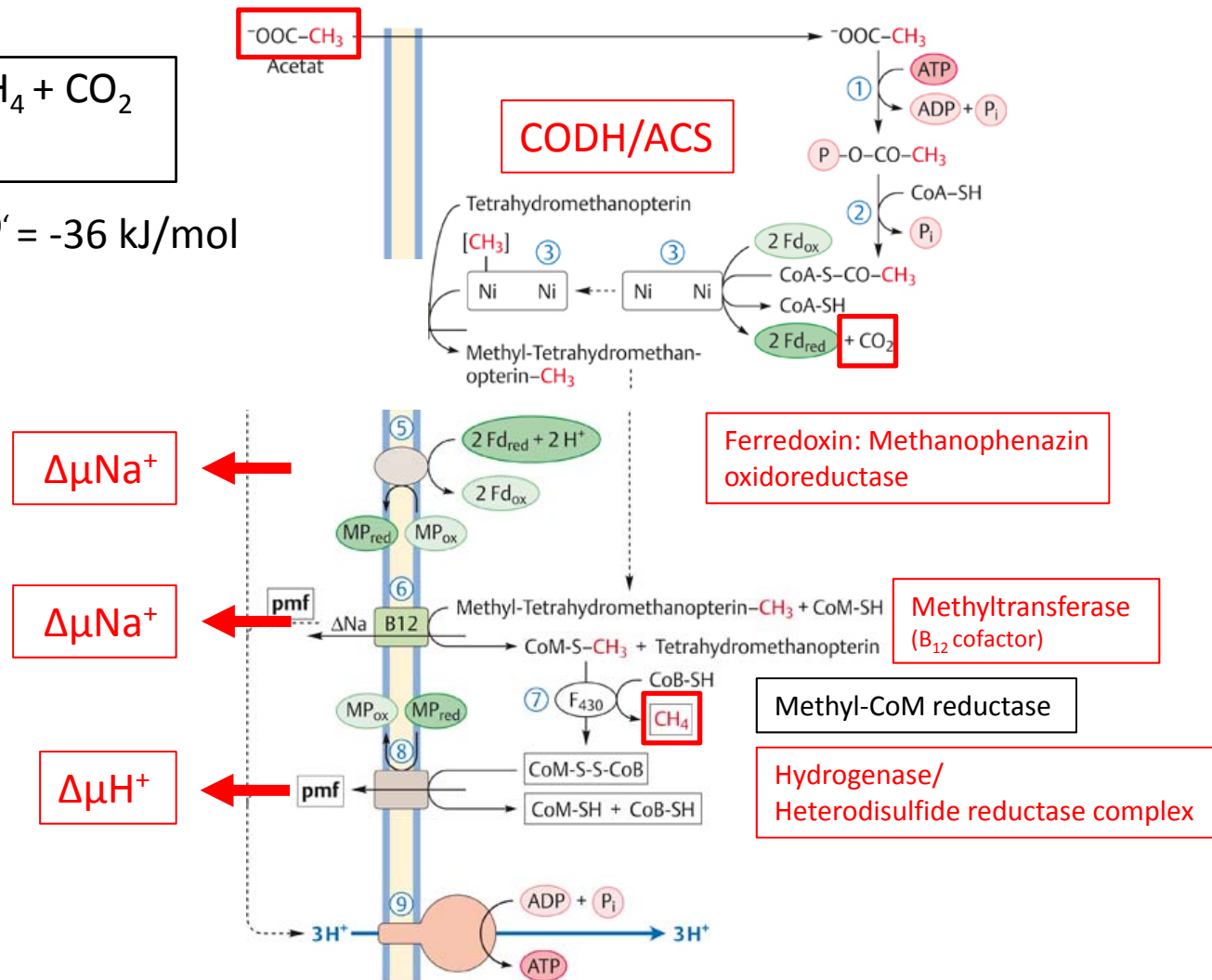
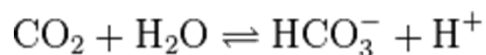
- Oxidation of the carboxyl group to  $\text{CO}_2$
- Reduction of the methyl group to  $\text{CH}_4$

Transport of 6-8  $\text{H}^+$  ( $\text{Na}^+$ )/acetate

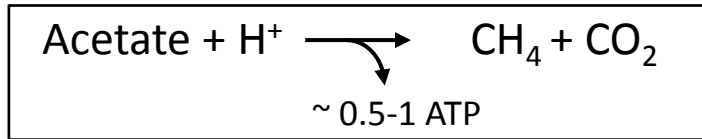
→ corresponding to 1,5 - 2 ATP

But: Activation of acetate to acetyl-CoA requires at least 1 ATP

Carbonic anhydrase:



# Methanogenesis from acetate



Disproportionation:  $\Delta G^{\circ} = -36 \text{ kJ/mol}$

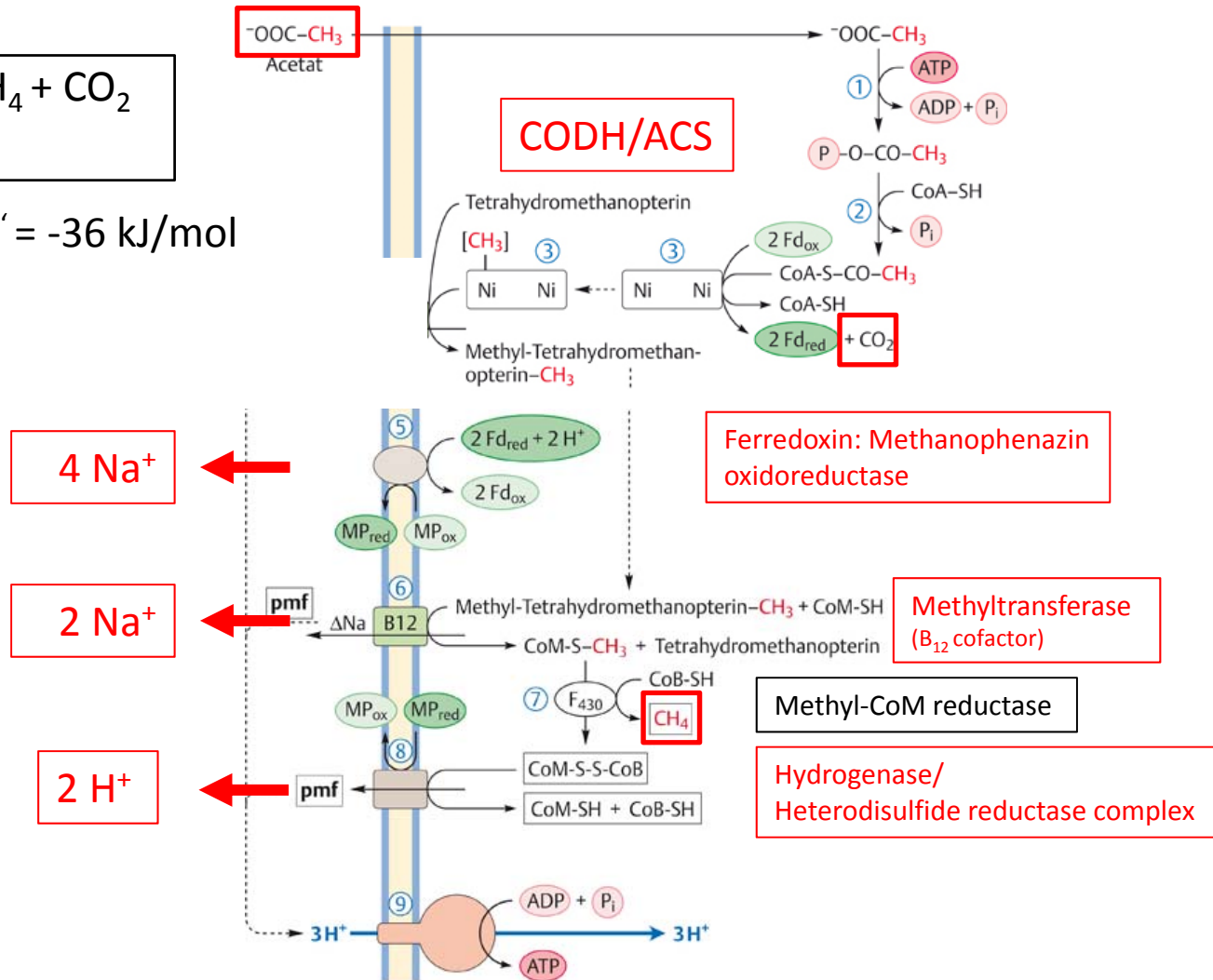
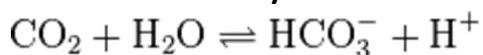
- Oxidation of the carboxyl group to  $\text{CO}_2$
- Reduction of the methyl group to  $\text{CH}_4$

Transport of 6-8  $\text{H}^+$  ( $\text{Na}^+$ )/acetate

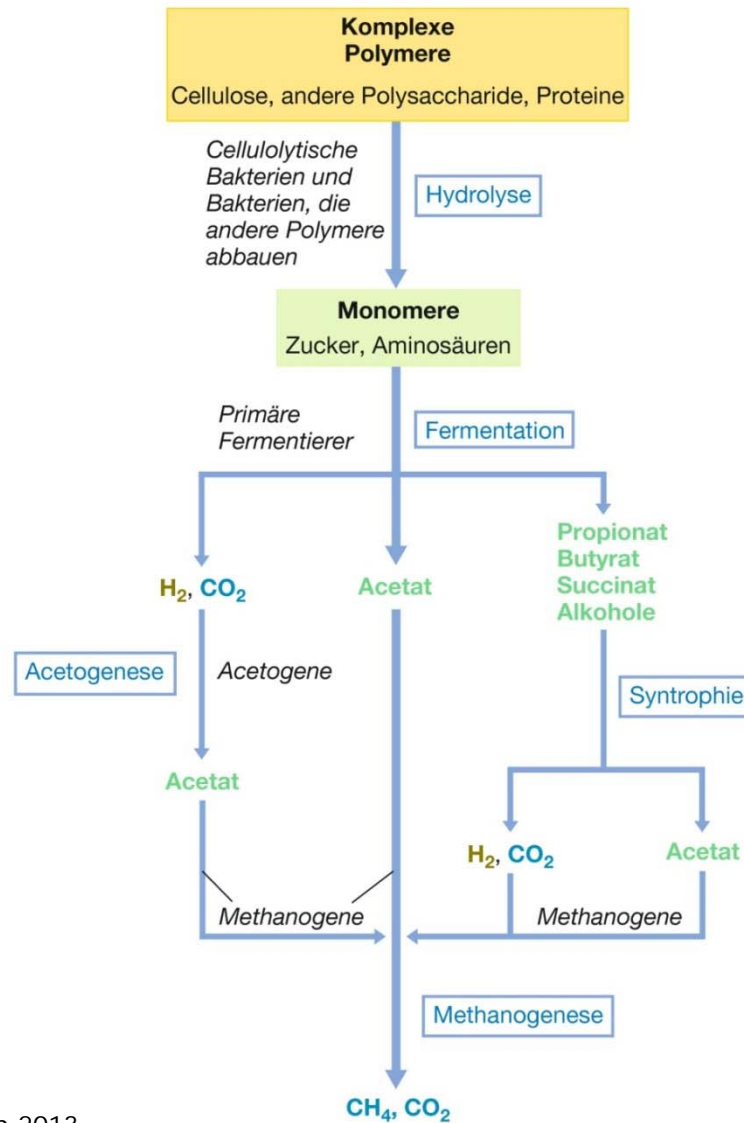
→ corresponding to 1,5 - 2 ATP

But: Activation of acetate to acetyl-CoA requires at least 1 ATP

Carbonic anhydrase:



# Anaerobic food chain

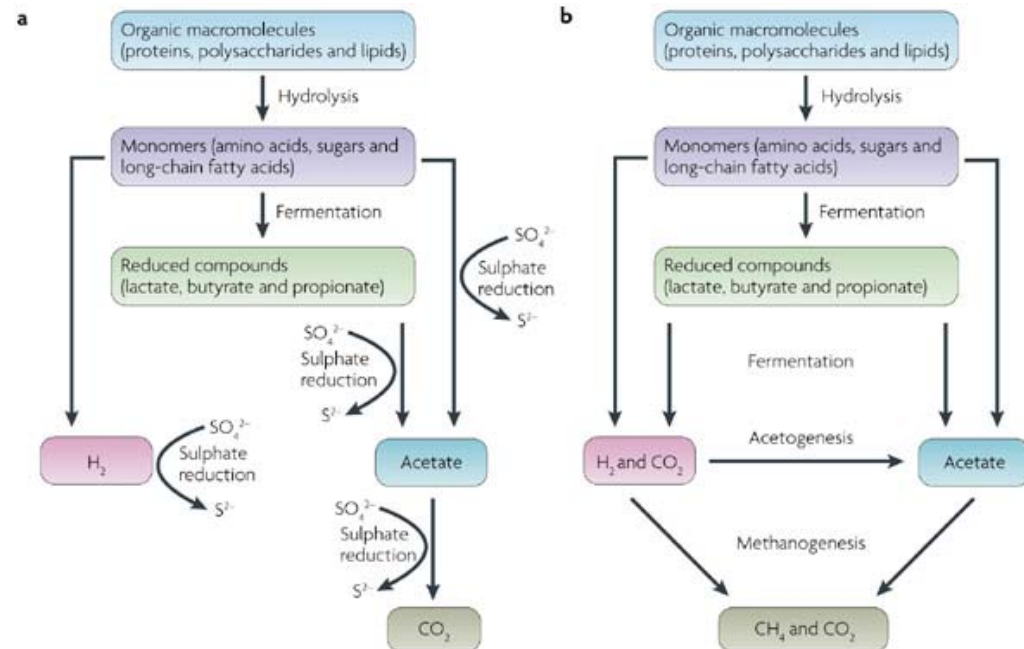


**Abbildung 24.5: Anoxischer Abbau.** Beim anoxischen Abbau kooperieren bei der Umwandlung komplexer organischer Substanzen von  $CH_4$  zu  $CO_2$  verschiedene Gruppen fermentativer Anaerobier. Diese Darstellung trifft auf Lebensräume zu, in denen sulfatreduzierende Bakterien eine untergeordnete Rolle spielen, zum Beispiel in den Sedimenten von Süßwasserseen, Klärschlammbioreaktoren oder dem Pansen.



# Sulfate reduction

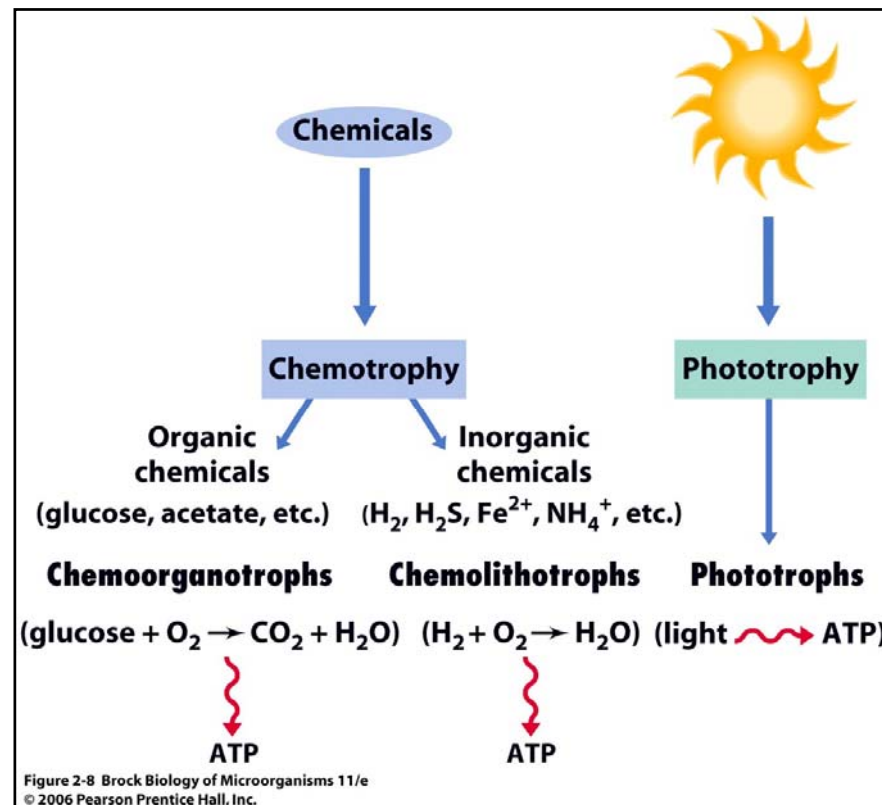
- Competition of sulfate reducers with syntrophic methanogenic communities and acetogens for fermentation products (lactate, propionate, acetate, H<sub>2</sub> in anaerobic environments)
  - Sulfate reducers with broader substrate spectrum than e.g. methanogens
  - Higher affinity and lower threshold for H<sub>2</sub>
- sulfate reducers outcompete methanogens at elevated SO<sub>4</sub><sup>2-</sup> concentrations e.g. in marine sediments



# Life style

<b>Energiequelle</b>	Licht	Photo-		
	Redoxreaktion	Chemo-		
<b>Elektronendon(at)or</b>	anorganischer Stoff		Litho-	
	organischer Stoff		Organo-	
<b>Kohlenstoffquelle</b>	anorganischer Stoff			Auto-
	organischer Stoff			Hetero-
				-trophie

- Microorganisms show a high metabolic diversity
- Play important roles in the biogeochemical cycles of elements

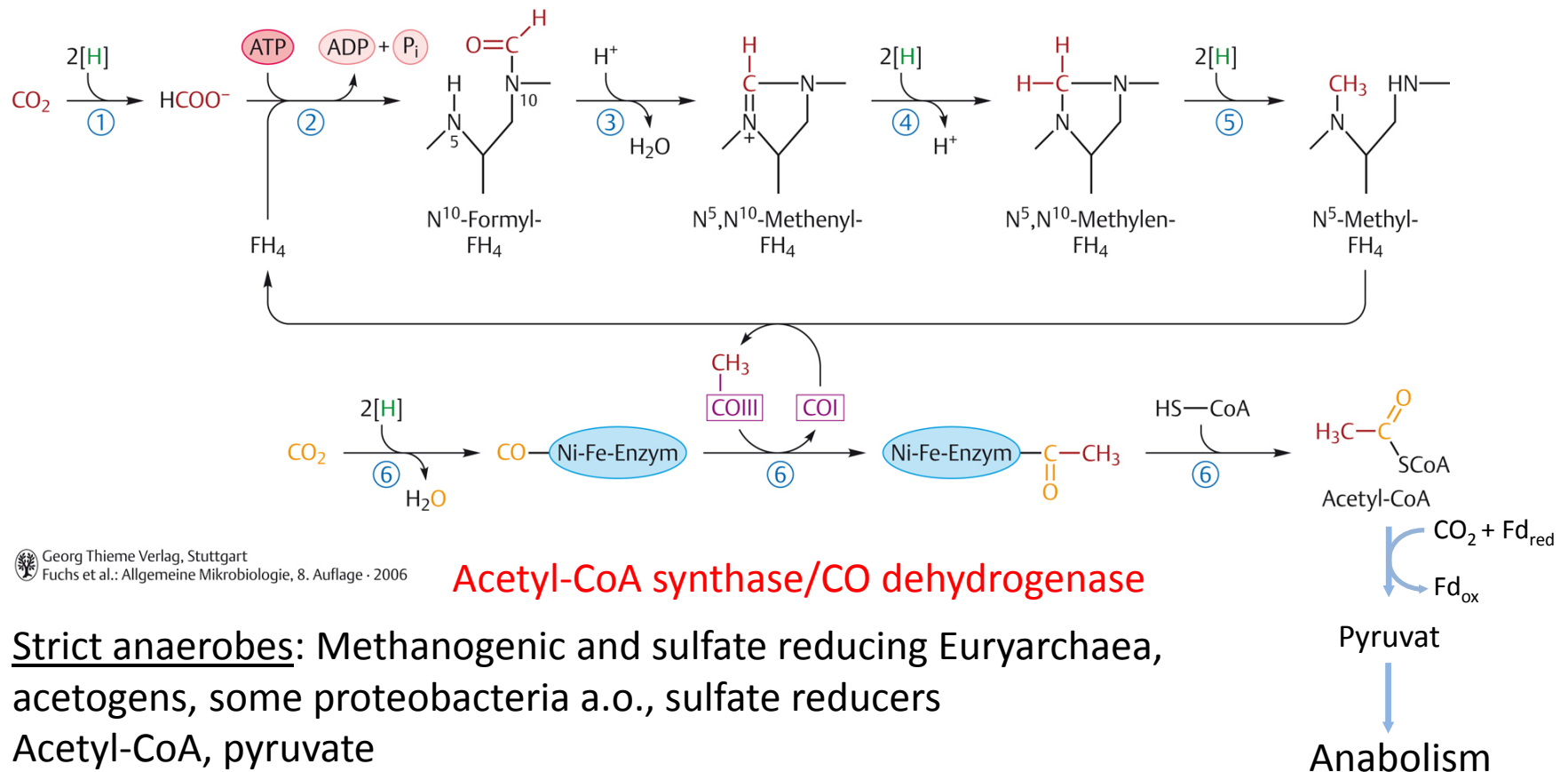
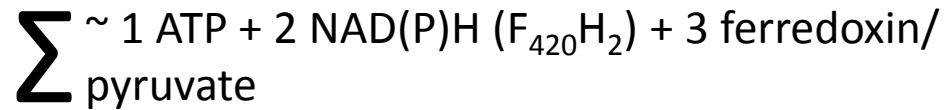


# CO<sub>2</sub> Fixation

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# Reductive acetyl-CoA pathway

reductive CODH/ACS pathway = Wood-Ljungdal pathway



Georg Thieme Verlag, Stuttgart  
Fuchs et al.: Allgemeine Mikrobiologie, 8. Auflage · 2006

## Acetyl-CoA synthase/CO dehydrogenase

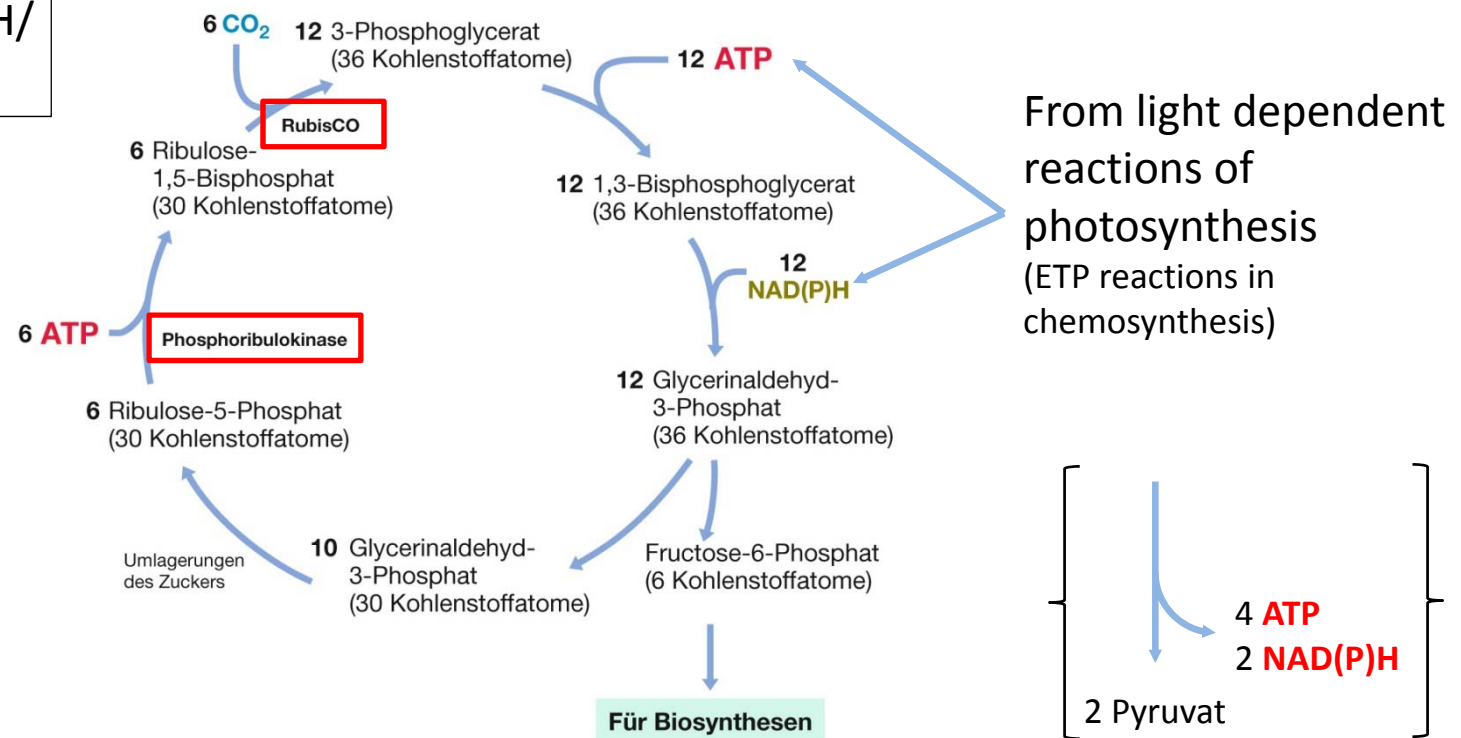
- Strict anaerobes: Methanogenic and sulfate reducing Euryarchaea, acetogens, some proteobacteria a.o., sulfate reducers
- Acetyl-CoA, pyruvate
- Well suited for the assimilation of C1 units

# Reductive pentose phosphate cycle

(Calvin-Benson cycle)

RubisCO = Ribulose-bisphosphate carboxylase/oxygenase

$\Sigma$  7ATP +5 NAD(P)H/  
pyruvate



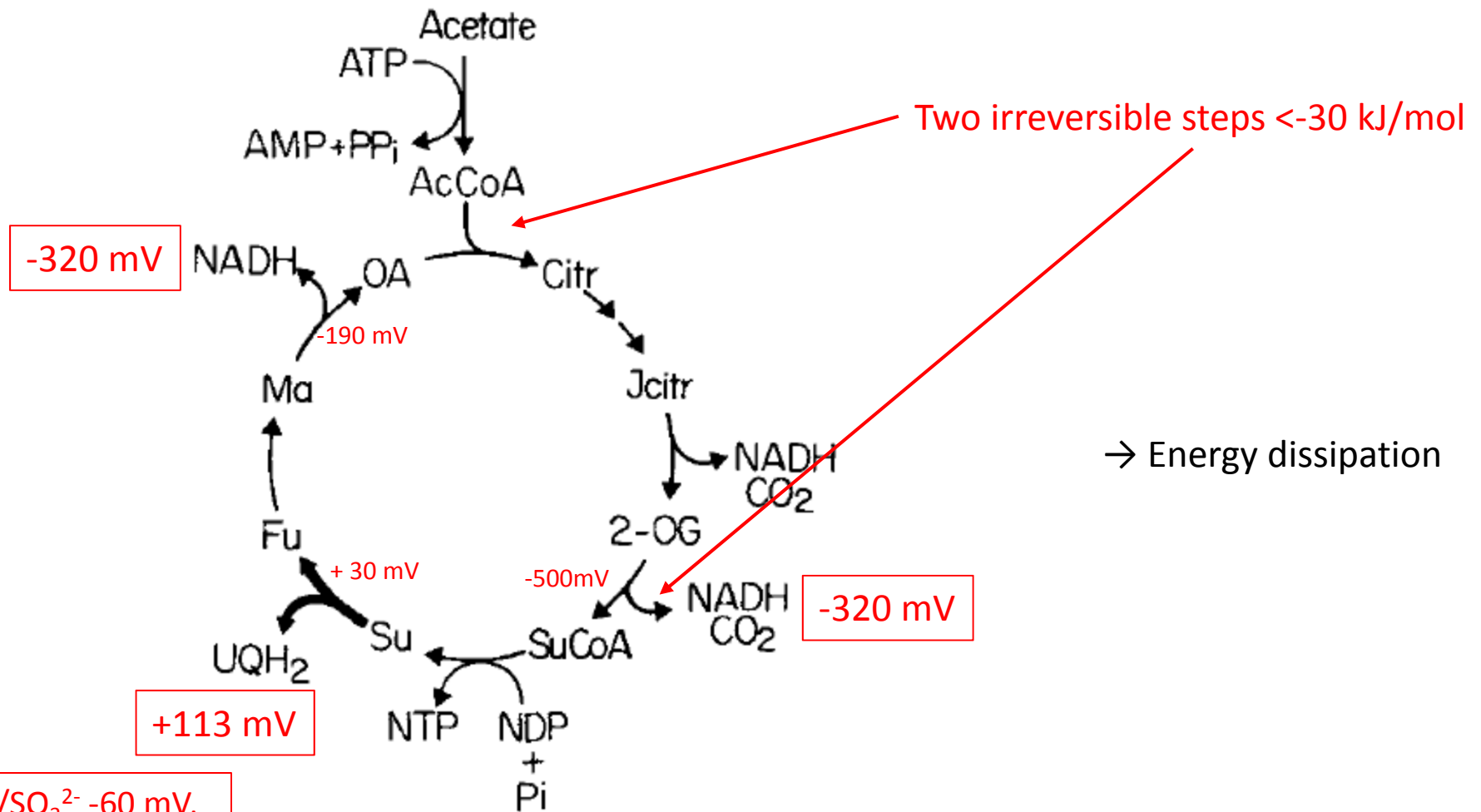
- Plants, algae, cyanobacteria, most aerobic and facultativ aerobic (chemolithoautotrophic) Bacteria, anoxygen phototrophic purple bacteria
- Triosephosphates, 3-phosphoglycerate, sugar phosphates as intermediates



# The Citric acid cycle (oxidative)

in aerobic organisms

Acetate activation (two ATP equivalents)



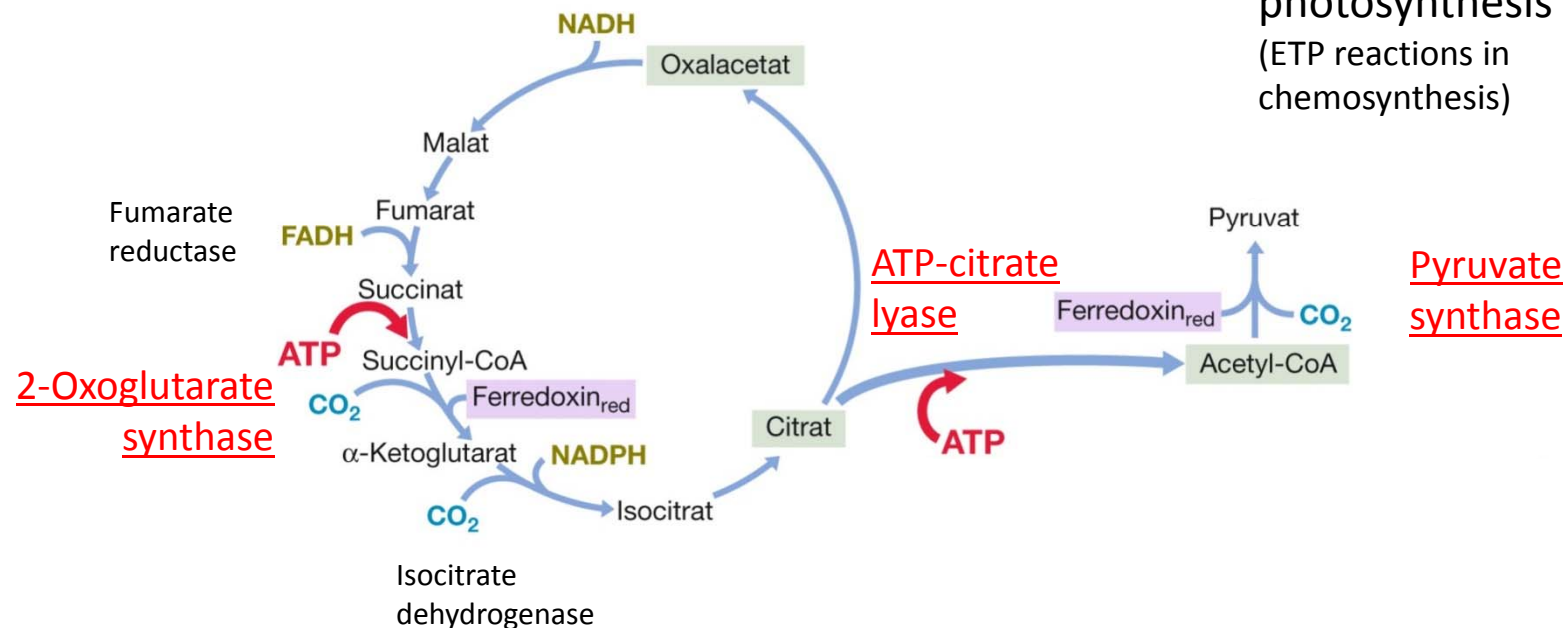
APS/SO<sub>3</sub><sup>2-</sup> -60 mV,  
SO<sub>3</sub><sup>2-</sup>/HS<sup>-</sup> -110 mV!

# Reductive citric acid cycle

(Arnon-Buchanan cycle)

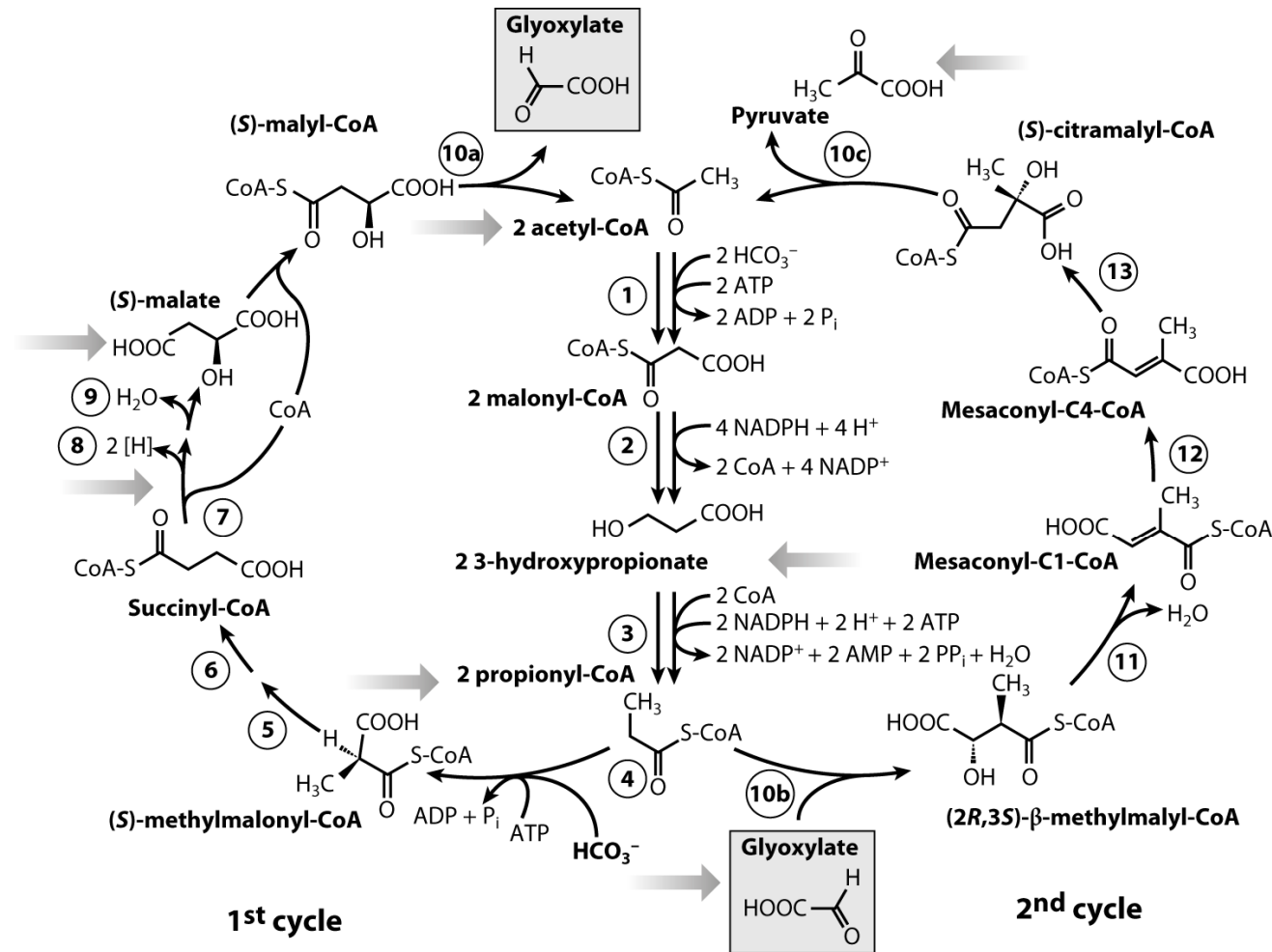
$\Sigma$  2 (-3) ATP + 3 NAD(P)H + 2 ferredoxin/  
pyruvate

ATP and ferredoxin from  
light dependent  
reactions of  
photosynthesis  
(ETP reactions in  
chemosynthesis)



- Anaerobic Green sulfur bacteria (Chlorobiales) and other Proteobacteria, Aquificales (microaerophilic)
- Acetyl-CoA, pyruvate, oxaloacetate, succinyl-CoA, 2-oxoglutarate, (PEP)
- Advantages under anaerobic, microaerophilic conditions

# Hydroxypropionate bi-cycle

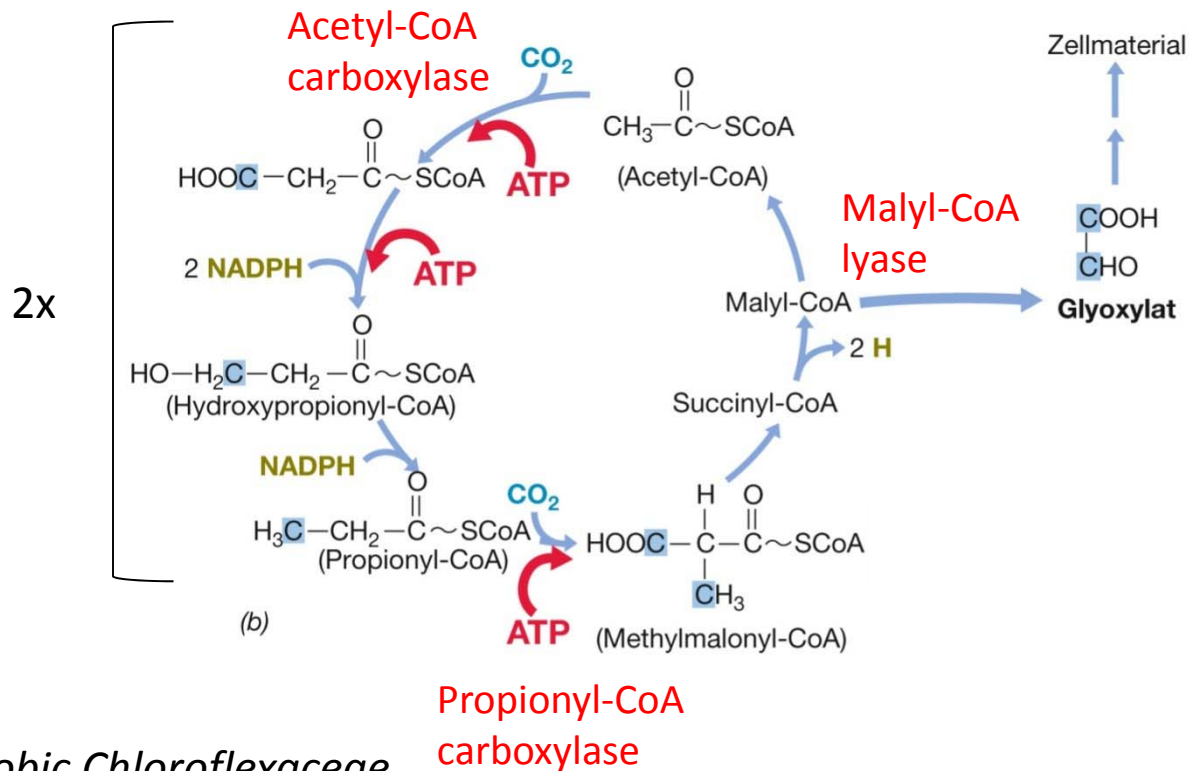


**AR** Fuchs G. 2011.  
Annu. Rev. Microbiol. 65:631–58

$\sum \sim 7 \text{ ATP} + 6 \text{ NAD(P)H} (-1\text{QH}_2) /$   
pyruvate

# 3-Hydroxypropionate bi-cycle

$\Sigma \sim 7 \text{ ATP} + 6 \text{ NAD(P)H} (- 1 \text{QH}_2) /$   
pyruvate

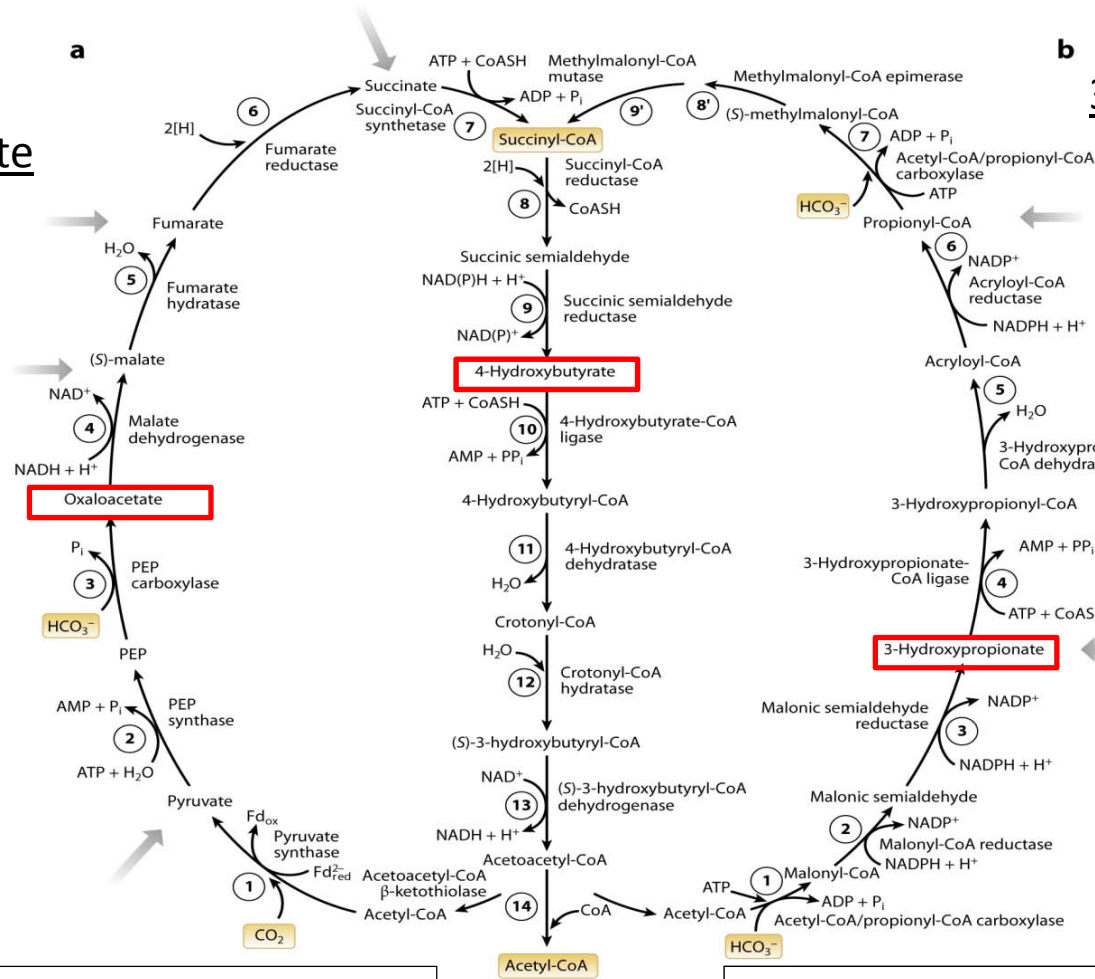


- *Anaerobic Chloroflexaceae*
- Acetyl-CoA, pyruvate, succinyl-CoA
- Suited for mixotrophic assimilation of fermentation products under oligotrophic conditions

# Dicarboxylate/4-Hydroxybutyrate cycle 3-Hydroxypropionate/4-Hydroxybutyrate cycle

## Dicarboxylate/ 4-Hydroxybutyrate cycle

Anaerobic  
Crenarchaeota  
(Thermoproteales,  
Desulfurococcales)



## 3-Hydroxypropionate/ 4-Hydroxybutyrate cycle

Aerobic  
Crenarchaeota (e.g.  
*Sulfolobus*) and  
Thaumarchaeota  
(Thermoproteales,  
Desulfurococcales)

$$\sum \sim 5 \text{ ATP} + 2 \text{ Fd}_{\text{red}} + \text{NAD(P)H} + 4 [\text{H}] / \text{pyruvate}$$

$$\sum \sim 6 \text{ ATP} + 3 \text{ NAD(P)H} + 4 [\text{H}] / \text{pyruvate}$$

# Questions 5

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- What is methanogenesis? What are the substrates, what the products?
- Which organisms perform methanogenesis?
- Name a special feature of methanogens/methanogenesis.
- How does methanogenesis from acetate basically works? What is oxidized, what is reduced? What is especially challenging with respect to energetics?
- How does chemolithotrophic methanogens fix CO<sub>2</sub>?
- Which are the energy gaining reactions in both types of methanogenesis?
- What is the lower pH<sub>2</sub> limit of methanogenesis and what does that mean for syntrophy?
- Name at least 3 pathways of CO<sub>2</sub> fixation!

# Fragen 5

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- Was ist Methanogenese? Was sind die Substrate, was die Produkte?
- Welche Organismen betreiben Methanogenese?
- Benennen Sie spezielle Eigenschaften der Methanogenen.
- Wie verläuft grundsätzlich die Methanogenese aus Acetat? Was wird oxidiert, was reduziert? Was ist im Hinblick auf die Energetik eine besondere Herausforderung?
- Wie fixieren chemolithoautotrophe Methanogene  $\text{CO}_2$ ?
- Welches sind die energieliefernden Reaktionen der in beiden Formen der Methanogenese?
- Was ist das untere Limit des  $\text{H}_2$  Partialdruckes für die Methanogenese und was bedeutet das für die Snytrophie
- Benennen Sie mindestens 3 Wege zur  $\text{CO}_2$  Fixierung



# Übung: Mikrobielles Wachstum I

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## Aufgabe:

1.) *E. coli* wurde auf eine  $OD_{600}$  von 0,1 aus einer exponentiell wachsenden Kultur auf das gleiche Medium (Minimalmedium mit 20 mM als C- & E-Quelle) überimpft und wächst exponentiell mit einer Verdopplungszeit von 40 min. Wie lange dauert es, bis die Kultur eine  $OD_{600}$  von 5,0 erreicht hat?

2.) Ihre Kurve soll in etwa alle 0.5 OD Einheiten einen Meßpunkt zeigen; zu welchen Zeitpunkten müssen Sie Probe nehmen?

# Übung: Mikrobielles Wachstum I

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## Aufgabe:

1. *E. coli* wurde auf eine  $OD_{600}$  von 0,1 aus einer exponentiell wachsenden Kultur auf das gleiche Medium (Minimalmedium mit 20 mM als C- & E-Quelle) überimpft und wächst exponentiell mit einer Verdopplungszeit von 40 min. Wie lange dauert es, bis die Kultur eine  $OD_{600}$  von 5,0 erreicht hat?

Wachstumsrate  $\mu$ :

$$\mu = \frac{\ln x - \ln x_0}{t - t_0}$$

Verdoppelungszeit  $t_D$ :

$$t_D = \frac{\ln 2}{\mu}$$

# Übung: Mikrobielles Wachstum I

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Wachstumsrate  $\mu$ :

$$\mu = \frac{\ln x - \ln x_0}{t - t_0}$$

Verdoppelungszeit  $t_D$ :

$$t_D = \frac{\ln 2}{\mu}$$

$$\rightarrow t - t_0 = \frac{\ln x - \ln x_0}{\mu}$$

$$\rightarrow \mu = \frac{\ln 2}{t_D} = \frac{0,69}{40 \text{ min}} = 0,017 \text{ min}^{-1}$$

$$\rightarrow t - t_0 = \frac{\ln 5 - \ln 0,1}{0,017 \text{ min}^{-1}} = \frac{1,61 - (-2,30)}{0,017 \text{ min}^{-1}} = \frac{3,91}{0,017 \text{ min}^{-1}}$$

$$\rightarrow t - t_0 = \frac{3,91}{0,017 \text{ min}^{-1}} = \underline{\underline{230 \text{ min}}}$$

# Übung: Mikrobielles Wachstum I

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Wachstumsrate  $\mu$ :

$$\mu = \frac{\ln x - \ln x_0}{t - t_0}$$

$$\rightarrow t - t_0 = \frac{\ln x - \ln x_0}{\mu}$$

$$\rightarrow t - t_0 = \frac{\ln 0,5 - \ln 0,1}{0,017 \text{ min}^{-1}} = \frac{-0,69 - (-2,30)}{0,017 \text{ min}^{-1}} = \frac{1,61}{0,017 \text{ min}^{-1}} = 95 \text{ min}$$

$$\rightarrow t - t_0 = \frac{\ln 1 - \ln 0,5}{0,017 \text{ min}^{-1}} = \frac{0 - (-0,69)}{0,017 \text{ min}^{-1}} = \frac{1,61}{0,017 \text{ min}^{-1}} = 40 \text{ min}$$

$$\rightarrow t - t_0 = \frac{\ln 1,5 - \ln 1}{0,017 \text{ min}^{-1}} = \frac{0,4 - 0}{0,017 \text{ min}^{-1}} = \frac{1,61}{0,017 \text{ min}^{-1}} = 23 \text{ min}$$

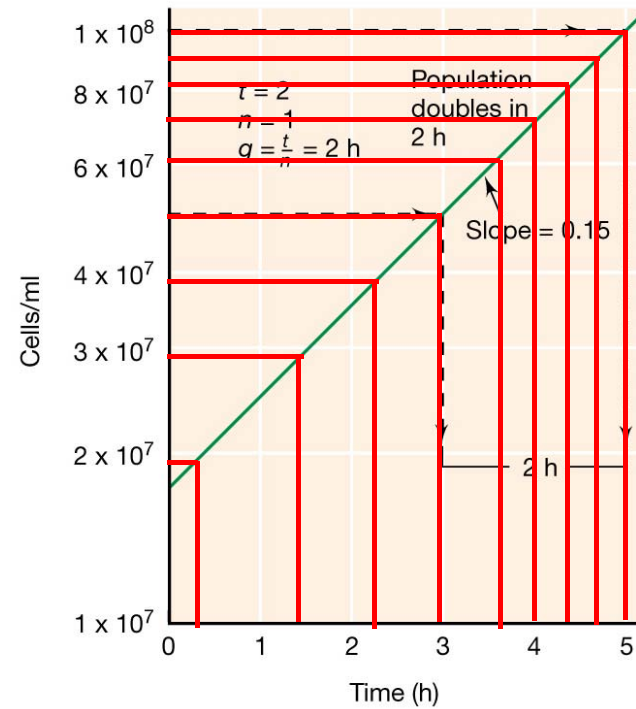
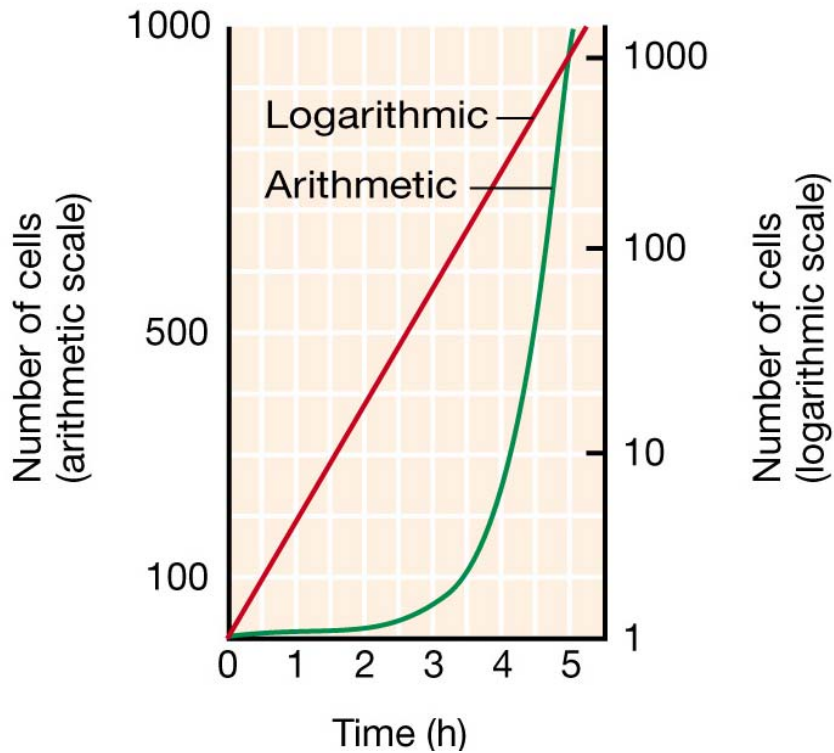
$\rightarrow 17,2 \text{ min} \rightarrow 13,1 \text{ min} \rightarrow \rightarrow \rightarrow \rightarrow 6,2 \text{ min}$  (OD 4.5 – OD 5)

Verdoppelungszeit  $t_D$ :

$$t_D = \frac{\ln 2}{\mu}$$

$$\rightarrow \mu = \frac{\ln 2}{t_D} = \frac{0,69}{40 \text{ min}} = 0,017 \text{ min}^{-1}$$

# Übung: Mikrobielles Wachstum I



# Übung: Mikrobielles Wachstum II

## Microbial growth and physiology: a call for better craftsmanship

Thomas Egli\*\*

*Environmental Microbiology, Swiss Federal Institute of Aquatic Science and Technology (Eawag), Dübendorf, Switzerland*

Virtually every microbiological experiment starts with the cultivation of microbes. Consequently, as originally pointed out by Monod (1949), handling microbial cultures is a fundamental methodology of microbiology and mastering different cultivation techniques should be part of every microbiologist's craftsmanship. This is particularly important for research in microbial physiology, as the composition and behavior of microbes is strongly dependent on their growth environment. It has been pointed out repeatedly by eminent microbiologists that we should give more attention to the media and culturing conditions. However, this is obviously not adhered to with sufficient rigor as mistakes in basic cultivation principles are frequently found in the published research literature. The most frequent mistakes are the use of inappropriate growth media and little or no control of the specific growth rate, and some examples will be discussed here in detail. Therefore, this is a call for better microbiological craftsmanship when cultivating microbial cultures for physiological experiments. This call is not only addressed to researchers but it is probably even more important for the teaching of our discipline.

**Keywords:** cultivation, batch, continuous culture, growth media, nutrient limitation, physiology

*"The study of the growth of bacterial cultures does not constitute a specialized subject or branch of research: it is the basic method of Microbiology"*

Monod (1949).

## Wie entwickelt man ein Medium?

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# Übung: Mikrobielles Wachstum II

Minimalmedium (definierte Zusammensetzung), C limitiertes Wachstum (heterotroph)

(A)

Elemental constituents in dry biomass	Average <sup>a</sup> (% of DW) <sup>b</sup>	Range (% of DW) <sup>b</sup>	Average $Y_{X/E}$ for C-limited growth
C	50	45 <sup>c</sup> -58 <sup>d</sup>	1
O	21	18 <sup>e</sup> -31 <sup>f</sup>	-
N	12	5 <sup>d</sup> -17 <sup>g</sup>	8
P	3	1.2 <sup>h</sup> -10 <sup>i</sup>	33
S	1	0.3-1.3	100
K	1	0.2 <sup>j</sup> -5 <sup>k</sup>	100
Mg	0.5	0.1 <sup>l</sup> -1.1	200
Fe	0.5	0.01-0.5	200

1 → 1 g TZ/ 1 g C

→ ~50% der C & E Quelle gehen in den Baustoffwechsel

Wie viel Stickstoff ( $\text{NH}_4\text{Cl}$  in mM) muss dem Medium mindestens zugegeben werden, damit der Stickstoff bei einer gegebenen C- & E-Quelle (20 mM Glucose) nicht limitierend wird?